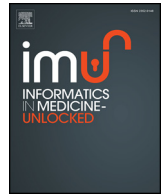




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## A comparative computational genomics of Ebola Virus Disease strains: *In-silico* Insight for Ebola control



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## ARTICLE INFO

## Keywords:

Comparative genomics

Ebola virus disease

*In-silico*

Insight

Ebola control

## ABSTRACT

Ebola Virus Disease (EVD), is a national epidemic in Countries affected. It is also a potential global public health pandemic. The menace of the disease outbreak among West and Central African nations, in recent years, has resulted in the death of many unsuspecting victims. The present study was conducted to present a systematic review of the literature, focusing on the control of Ebola Virus Disease (EVD), among human subjects. It also centered on a bioinformatics analysis of five different strains of Ebola virus.

Research articles published between 2008 and 2018, on EVD control studies, were systematically reviewed. Four online databases were searched for the purpose of this review. These include: Science Direct, Google Scholar, SpringerLink and PubMed. Study outcomes were extracted. The outcomes were summarized and categorized. Five different strains of Ebola virus were obtained from the NCBI database, specifically the Entrez Genome database. Bioinformatics analysis was performed using Muscle software, RawXL, Treview, iTOL and Clustal X. Bioinformatics analysis was performed on five selected strains of Ebola virus (Reston, Bundingbugyo, Zaire, Sudan and Tai forest). Evaluation of the phylogenetic tree was performed by using MEGA X and PHYLIP software.

237,498 publications were identified, out of which 104 research articles, from different regions of the world, fulfilled our inclusion criteria. Insight was gained for the control of EVD from these studies. Of the studies reviewed, 23 articles focused on vaccines/vaccination-related Ebola control research, 12 studies on modeling and simulation-related Ebola control research, 41 on drugs and therapeutics-related Ebola control research, and 28 focused on other experimental studies (such as biological experiments, bioinformatics experiments, travel border control measures, educational campaign measures, hand and environmental sanitization, amongst others). Very few modeling and simulation studies have been conducted on the control of EVD in the last 10 years. Thus, there is the need for more modeling and simulation-related ebola control research. Comparative computational genomics of the five Ebola virus strains produced phylogenetic trees in different shapes. An evaluation of the phylogenetic tree was performed. Results showed that *Taiforest Ebola virus* and *Bundibugyo Ebola virus* are closely related. The results also revealed that *Sudan and Reston Ebola virus* are closely related. *Zaire Ebola virus* stood out from all the others. It may be possible to adopt similar Ebola control measures against Ebola virus strains that are closely related.

Insight from these results, can facilitate the development and production of multi-protective, multi-treatment drugs, multi-protective vaccines and antivirals, against these ebola virus disease strains.

The results of the evaluations of the phylogenetic tree can be assistive in providing insight into the origin, evolution, and possible structural and genetic mutations of the Ebola virus. It can also provide insight for inferring the structural and functional properties of each Ebola virus. The knowledge of such inference can be useful for EVD control. This can bring about a radical transformation in control efforts for disease.

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<https://doi.org/10.1016/j.imu.2018.07.004>

Received 3 January 2018; Received in revised form 28 June 2018; Accepted 17 July 2018

Available online 23 July 2018

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## 1. Introduction

Ebola Virus Disease (EVD, henceforth), is a national epidemic in affected countries. It is also a potential global public health pandemic. Particularly disturbing, is the resurgence and re-emergence of the disease in some African countries. The menace of the disease outbreak among West and Central African nations, in recent years, has resulted in the death of many unsuspecting victims [1,2,3,4]. In 2014, the spate of EVD in West African countries spurred untold hardships, many cases of fatalities, discrimination, stigmatization, isolation, and mockery of affected countries [5,6]. After curtailing the disease, a few years later, EVD re-emerged and it was confirmed in DR Congo on the 11th of May 2017. The disease killed some inhabitants of a remote village in the Northeastern region of DR Congo. It has been reported that delayed diagnosis and lack of prompt reporting of new cases of EVD affects control efforts, especially in rural areas [2]. Similarly, in a recent development, the DR Congo government recently confirmed a new outbreak of EVD on the 8th of May 2018. There were two 2 confirmed cases of EVD incidence [7]. Some of the pertinent questions that are readily evinced are: (i) what are the various efforts in Ebola research that focus on controlling EVD transmissions among human subjects? (ii) What kind of research has been conducted, that can provide insight or *in-silico* analysis for control of Ebola? (iii) How much of each kind of EVD control research has been conducted in the last decade? (iv) Is it possible to gain insight on the control of EVD from the bioinformatics analysis of different Ebola virus strains?

In order to address these pertinent questions, our study focused on a 10-year systematic review of previous studies, which centered on the control of EVD. Our study was conducted with a view to obtain insight in the control of the disease. We performed bioinformatics analysis (Multiple Sequence Alignment (MSA) and phylogenetic analysis) on five different strains of Ebola virus. We also evaluated the results of the bioinformatics analysis.

The aim of this study was to conduct a comprehensive systematic review on investigations that have determined potential control measures against EVD transmission. It was also to perform a bioinformatics analysis on five different strains of Ebola virus. The significance of performing bioinformatics analysis was to understand the relationship (s) between the five different strains of Ebola virus. It was also to gain insight toward the control and prevention of the spread of the disease. The paper is structured as follows: firstly there is a methodology of the literature review, then a bioinformatics analyses, results, discussion, recommendations and conclusion.

## 2. Methodology of literature review

### 2.1. Strategy of searching

Databases of the scientific literature such as ScienceDirect, SpringerLink, PubMed, and Google Scholar, were searched between the period of January 1st, 2008 and February 28th, 2018. Relevant subject heading and keywords were used. The databases, search terms, and number of articles found, are provided in Table 1. Multiple keywords were chosen, based on their respective relevance to identify studies on 'Ebola control', 'reduction of Ebola transmissions', 'bioinformatics analyses', 'phylogenetic analyses', 'comparative genomics', 'control'. The combination of the keywords, was used for conducting search through each scientific database. Scientific articles that relate to empirical studies, only on humans (human subjects), and published in the English language, were considered for the systematic review.

### 2.2. Selection and exclusion criteria of study

The initial search resulted in the identification of 237,498 articles (See Fig. 1 and Table 1). The next stage was the title review stage. This was conducted by adopting the inclusion criteria. 234,830 articles were

**Table 1**  
Databases, number of articles synthesized and integrated to the system.

s/n	Scientific Databases	Search terms	Initial Results	Articles Synthesized & Integrated
1	Science Direct Database <a href="https://www.sciencedirect.com/">https://www.sciencedirect.com/</a>	(multiple sequence alignment* OR 'Ebola virus') OR (Phylogenetic analyses* OR 'Ebola virus') OR (Comparative genomics* OR 'Ebola virus') OR (Bioinformatics analysis* OR 'Ebola virus') AND (Ebola control) OR (Ebola transmission) OR (reduction)	6000 articles	6
2	Google Scholar <a href="https://scholar.google.com">https://scholar.google.com</a>	(multiple sequence alignment* OR 'Ebola virus') OR (Phylogenetic analyses* OR 'Ebola virus') OR (Comparative genomics* OR 'Ebola virus') OR (Bioinformatics analysis* OR 'Ebola virus') AND (Ebola control) OR (Ebola transmission) OR (reduction)	940 articles	12
3	Springer Link <a href="https://link.springer.com/">https://link.springer.com/</a>	(multiple sequence alignment* OR 'Ebola virus') OR (Phylogenetic analyses* OR 'Ebola virus') OR (Comparative genomics* OR 'Ebola virus') OR (Bioinformatics analysis* OR 'Ebola virus') AND (Ebola control) OR (Ebola transmission) OR (reduction)	950 articles	6
4	PubMed [MEDLINE DATABASE] PubMed <a href="https://www.ncbi.nlm.nih.gov/pubmed/">https://www.ncbi.nlm.nih.gov/pubmed/</a>	(multiple sequence alignment* OR 'Ebola virus') OR (Phylogenetic analyses* OR 'Ebola virus') OR (Comparative genomics* OR 'Ebola virus') OR (Bioinformatics analysis* OR 'Ebola virus') AND (Ebola control) OR (Ebola transmission) OR (reduction)	229,608 articles	80
			Total Articles Reviewed =	237,498
				104

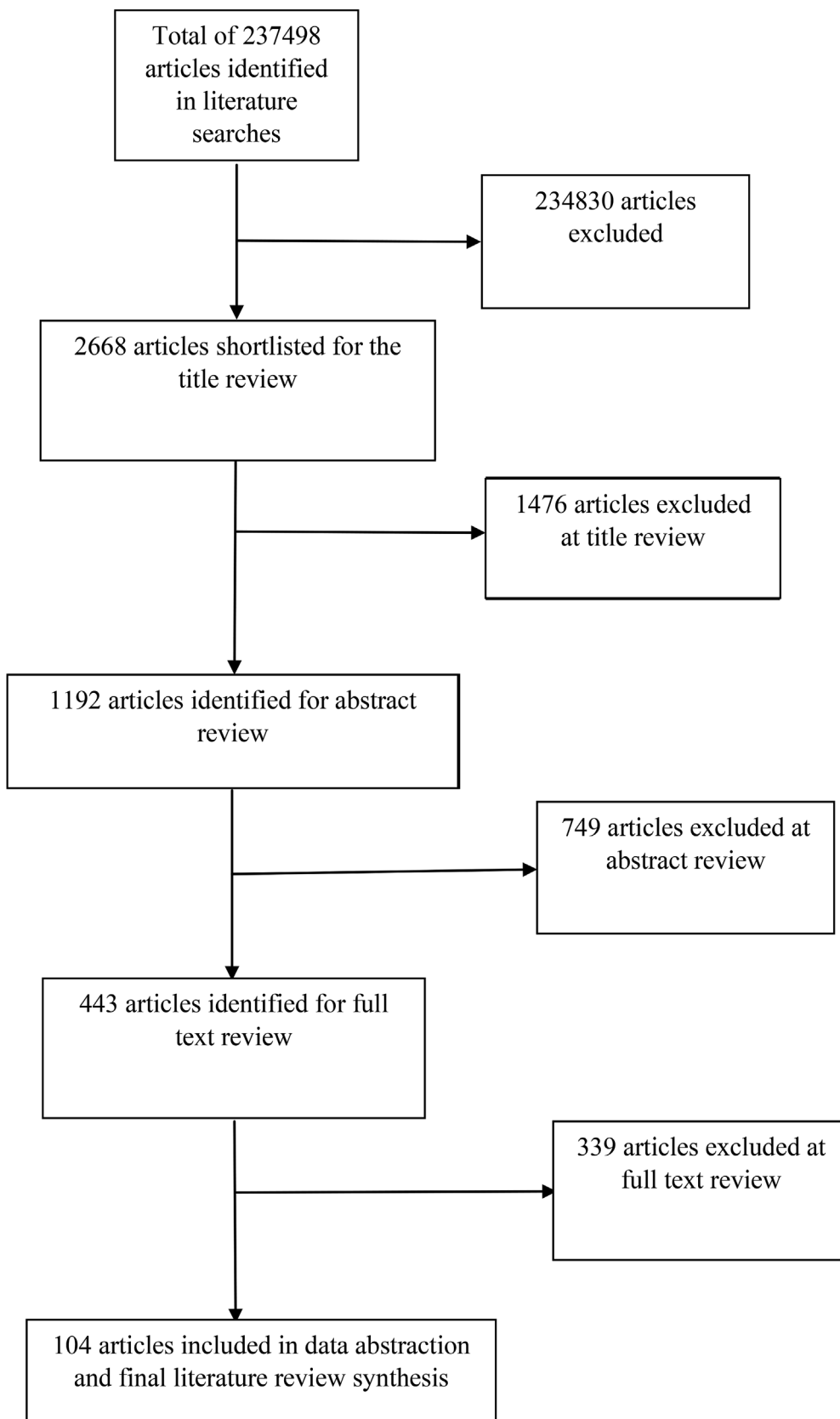


Fig. 1. The literature review process with the number of studies identified, excluded and included.

excluded after specifically reviewing each of the articles for the title review. At the title review stage, 2668 articles were shortlisted. The next stage was the abstract review stage. 1476 articles were excluded.

At the abstract review stage, 1192 articles were identified for abstract reviews. Following the exclusion of 749 articles based on prior specified inclusion criteria, the full text of 443 articles were reviewed and studied

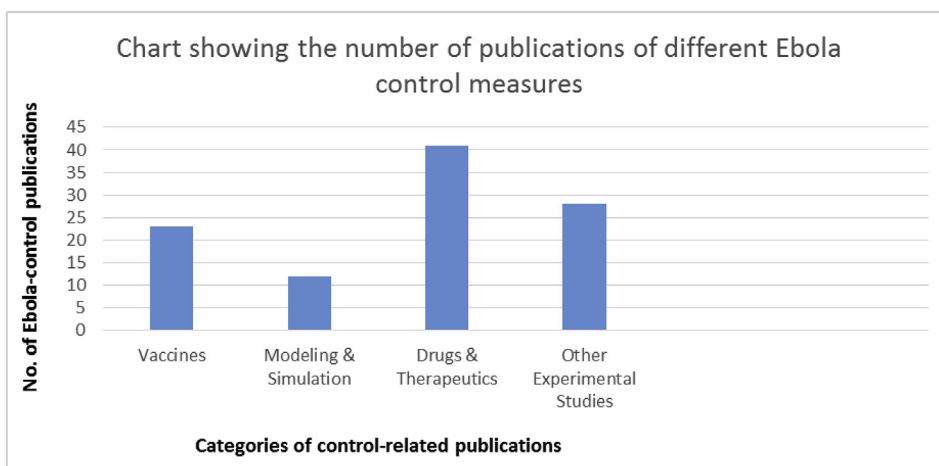


Fig. 2. Bar charts showing the classification of synthesized EVD control studies into Potential Control/intervention groups.

in details. Subsequently, 339 articles were excluded from the review. Finally, 104 articles met the inclusion criteria. These 104 articles were integrated and synthesized into the review [Reference [8–50][51–90] [91–112]] (See Fig. 1 and Supporting material 1).

2.3. Extraction of data and synthesis

For each paper reviewed that met our inclusion criteria, the following data were extracted: Authors, year, study method, study design, study site/region/country, data used/data collection methods/specimen/population involved/type of participants, key discoveries, title/issue/product investigated and *in-silico* insight/insight derived for EVD control. Results, titles, key discoveries and study methods were stated or in some cases paraphrased (see supporting material 1).

2.4. Bioinformatics analysis

The purpose of the bioinformatics analysis was to understand the relationships among the five Ebola virus strains. It was also to gain insight into control measures against EVD. The updated versions of the completely sequenced genomes of Ebola virus, was obtained from the NCBI database. Five different strains of Ebola virus genome data were obtained from the source: <https://www.ncbi.nlm.nih.gov/genomes/GenomesGroup.cgi?taxid=186536>. The authors obtained the sequences from the Entrez Genome database. The summary of the data is shown in Table 2. The Ebola virus genome data were obtained from

isolates of Ebola virus from Zaire (now DR Congo), Tai Forest (in the present Cote d’ivoire), Sudan, Reston (in the USA), and Bundibugyo (a town in Western Uganda). A bioinformatics analysis (multiple sequence alignment and phylogenetic analyses) was conducted.

Complete genomes of five Ebola virus isolates were downloaded from the NCBI website. The *Bundibugyo Ebola virus* genome data has accession number NC\_014373; the *Reston Ebola virus* updated genome data has an accession number NC\_004161; the *Sudan Ebola virus* genome data is a recently updated data with accession number NC\_006432; the *Tai Forest Ebola virus* updated genome data has accession NC\_004161. Finally, the *Zaire Ebola virus* has an accession number of NC\_002549. More information about these data can be found in Table 2.

2.5. Multiple sequence alignment and phylogenetic analyses

During analysis, the complete genome of Ebola virus isolates were downloaded. The genomes were in the FASTA file format (genomes\_e-bov.fasta). These are respectively *Bundibugyo Ebola virus* complete genomes, *Reston Ebola virus* complete genomes, *Sudan Ebola virus* complete genomes, *Tai forest Ebola virus* complete genomes, and *Zaire Ebola virus* complete genomes. These five *Ebola virus* species are cRNA molecules. This means that they are RNA viruses, with a length of about 19,000 nucleotides each. A Multiple Sequence Alignment (MSA) of all Ebola genomes was performed, using Muscle version 3.8.3.1 [113,114]. An aligned FASTA file format was created with it (genomes\_Ebola\_5.fasta). A

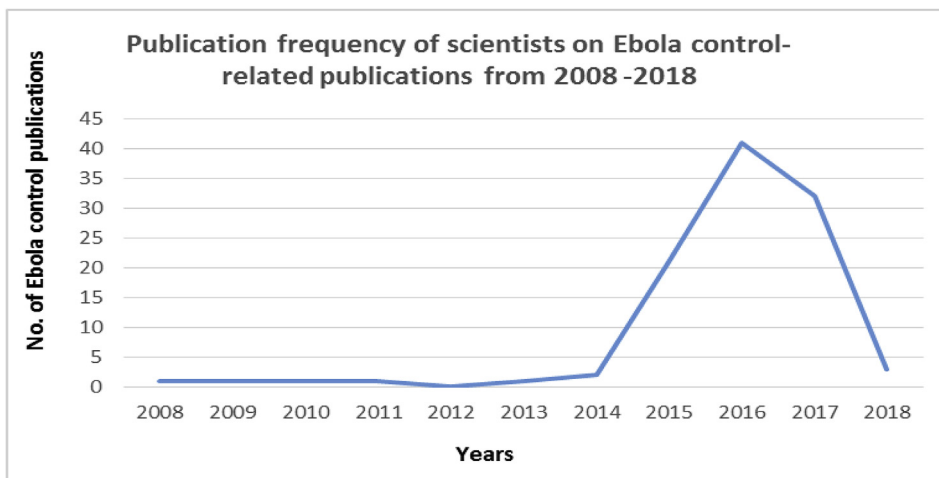


Fig. 3. Graph depicting the publication frequency of scientists on EVD control-related publications from 2008 to 2018.

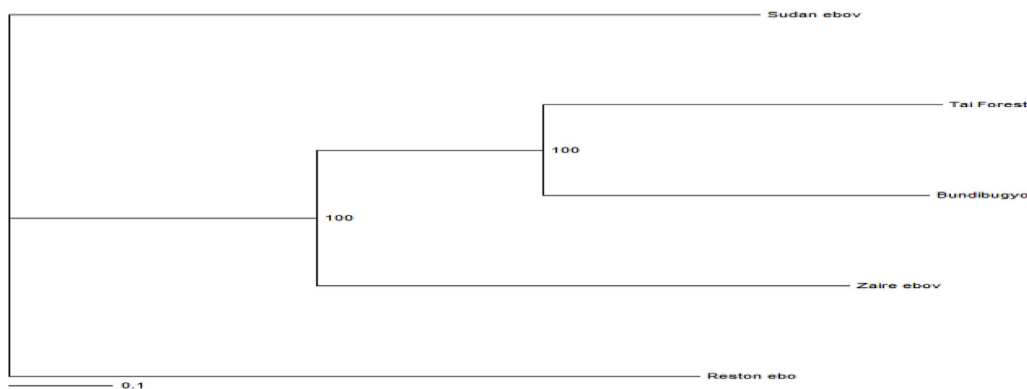


Fig. 4. Phylogram depicting the relationships among the five different strains of Ebola virus.

visualization tool, Seaview4 [115], was used to view the aligned output. Clustal X [116], was utilized to convert our aligned fasta file into phylip format. This contains 5 sequences and 20,308 base pairs.

RAxML [117] was used for the tree building and drawing process. The tree-building technique uses the Maximum Likelihood (ML, henceforth) method to build trees. RAxML [117], was utilized to perform a ML search, based on the genome sequences, and then bootstrapped 20 times. ML is a statistical technique of finding the value of one or more parameters, for given statistics, which makes the known likelihood distribution maximum [123]. Bootstrapping is another statistical technique that assists in providing allowance for assigning measures of accuracy [124]. It relies on random sampling with replacement [125,126]. The genome evolution was then modeled and the Phylip file was used to generate a newick file that produced the phylogenetic tree. A Tree Viewer [118,119,and120] was then used to view the tree. Treeview X [ [118,119,and 120]], was mostly used for generating the trees in different formats (the slanted cladogram, rectangular cladogram, and the phylogram). These trees were confirmed with iTOL [119]. Here is the tree generated in iTOL: <http://itol.embl.de/tree/100366160282901527527043>.

2.6. Evaluating the phylogenetic tree

Evaluating phylogenetic trees is significant because it helps to reveal how reliable the tree is. There are several methods, studies and tools for evaluating phylogenetic trees. Some of them include phylogenetic tree evaluation tools and studies, such as can be found in the following literature [127–157,161].

Two evaluations were performed on the phylogenetic tree. In order

to conduct a bootstrap (or jackknife, or permutation test), with some method in the PHYLIP package [158], these programs were used: Seqboot, Dnapars and Consense.

First, Seqboot was run to accept the original Ebola genome sequence data set, and produce a large number of bootstrapped or jackknifed data sets (with range between 100 and 1000). In our study, a value of 100 was selected.

Next, the phylogeny estimate for each of these was determined. This estimates were calculated according to the particular method of interest. For instance, to use the DNA parsimony algorithm, we first ran Seqboot, and generated a file with 100 bootstrapped data sets. The phylogeny estimate for each of the Ebola viruses was determined using the DNA parsimony algorithm. The Bundingbugyo Ebola virus was used as the outgroup. One hundred replicates were utilized for the phylip bootstrapping algorithm. A random number seed of three was employed.

The file produced was fed as input file into the Dnapars. The Dnapars was run with the Multiple Data Sets option, and informing it to expect 100 data sets.

The output from this analysis produced a large output file, as well as a tree file, with the trees from the 100 data sets. The tree file later served as the input for the Consense tree.

For DNA parsimony, 10000 number of trees were used to save. The input order of sequences were randomized using a seed of 13, and performed 10 times.

When Consense is run, this resulted in the majority rule consensus tree. This produced the outcome of the analysis: genomes\_Ebola\_final\_phylip\_output(1) and genomes\_Ebola\_final\_phylip\_output\_tree (1). The evaluation results from this exercise are depicted in

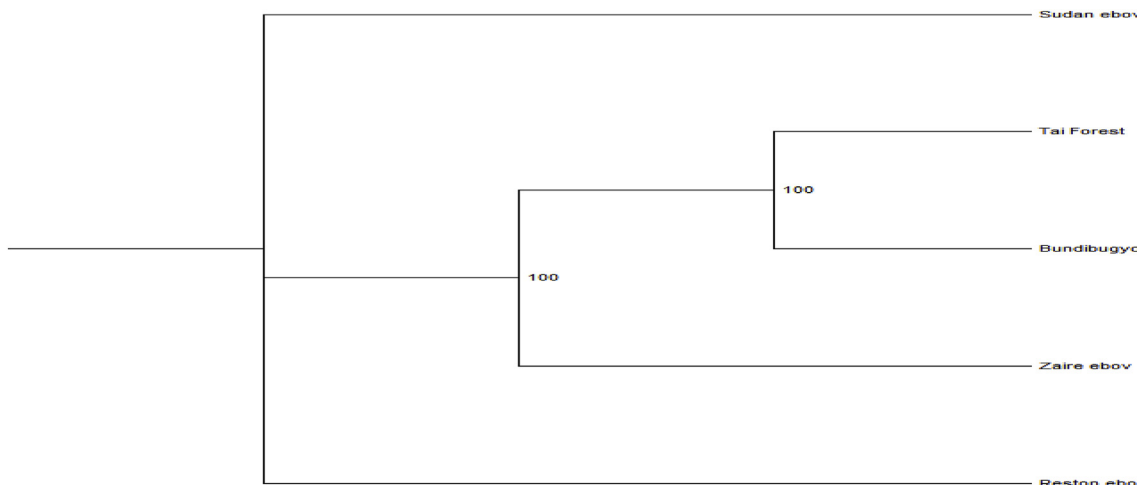


Fig. 5. Rectangular cladogram depicting the relationships among the five different strains of Ebola virus.

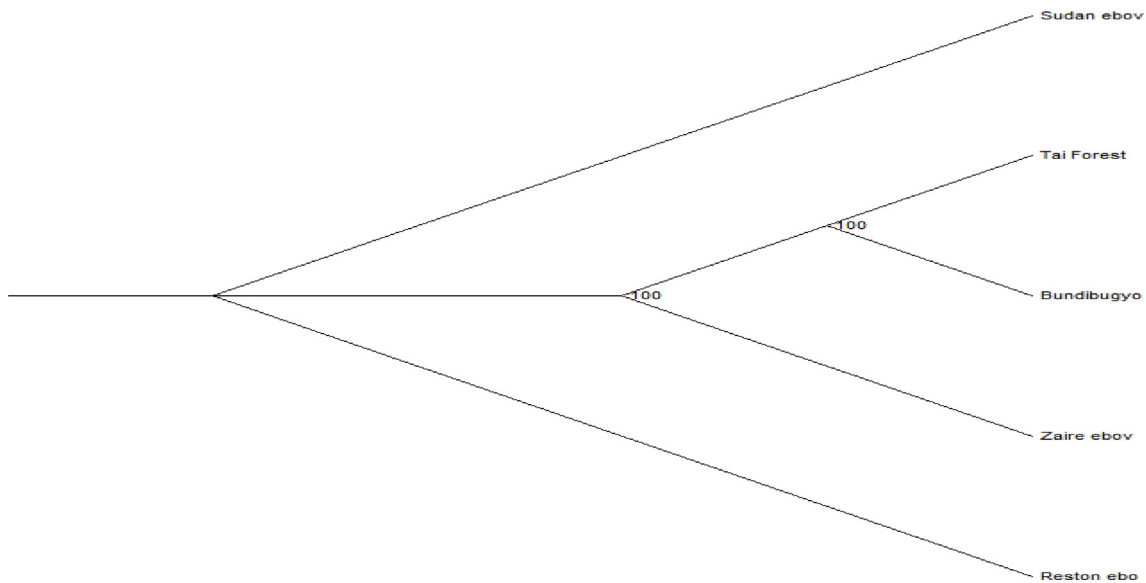


Fig. 6. Slanted cladogram depicting the relationships among the five Ebola virus strains.

Figs. 12 and 13. Online Tree viewer, ETE Toolkit phylogenetic tree viewer [160], was incorporated to visualize the results.

A second evaluation of the phylogenetic tree was then performed. For the second evaluation of the phylogenetic tree, MEGA X [159] and Clustal X [116], were used. Clustal X was used for bootstrapping, by incorporating the Neighbor Joining Clustering algorithm, and then a random number generator seed of 100 was used. The number of bootstrap trials was 1000.

MEGA X was employed to generate the newick tree. The newick tree was then viewed in different styles, as shown in Figs. 8–11.

### 3. Results

237,498 publications were identified. Out of these publications, 104 studies fulfilled our inclusion criteria. The studies were heterogeneous in the aspect of the study method, key discoveries, and insight gained towards control of Ebola, amongst others (see supporting material 1). The publications that fulfilled the inclusion criteria were classified into different groups - see Table 3 and Fig. 2, for the classification. The studies included were classified according to the EVD control intervention conducted. The interventions were classified into: drug & therapeutics, vaccines, modeling & simulations, and other experiments (See Table 3 and Fig. 2). Of the studies reviewed, 41 were drugs and therapeutics-related Ebola control research, 23 research publications were Ebola control vaccine-related studies, and 12 publications were EVD-control research that adopted a modeling and simulation approach [8–112]. Finally, 28 publications fell into the category of other experiments [8–112]. Other experiments include: biological experiments, bioinformatics experiments, border control measures, educational campaign measures, hand sanitation, and environmental sanitization, amongst others.

Furthermore, the trend and numbers of Ebola-control research from 2008 until 2018 were monitored. It was evident that from 2008 to 2013, there were very few publications. In 2014, the number of publications increased to 2 [87,96]. The number of publications of Ebola control-related research outputs in 2015, 2016, 2017 and 2018, were respectively, 21 research publications [60,67–80,83–85,89,93,104], 41 research publications [35–59,61–64,66,82,86,90,91,100,106–110,112], 32 research publications [8,10–34,81,88,92,94,99,105] and three research publications [9,101,103]. The results of the publishing frequency and trend, for scientists worldwide, are depicted in Table 4 and Fig. 3.

#### 3.1. Phylogenetic analysis result

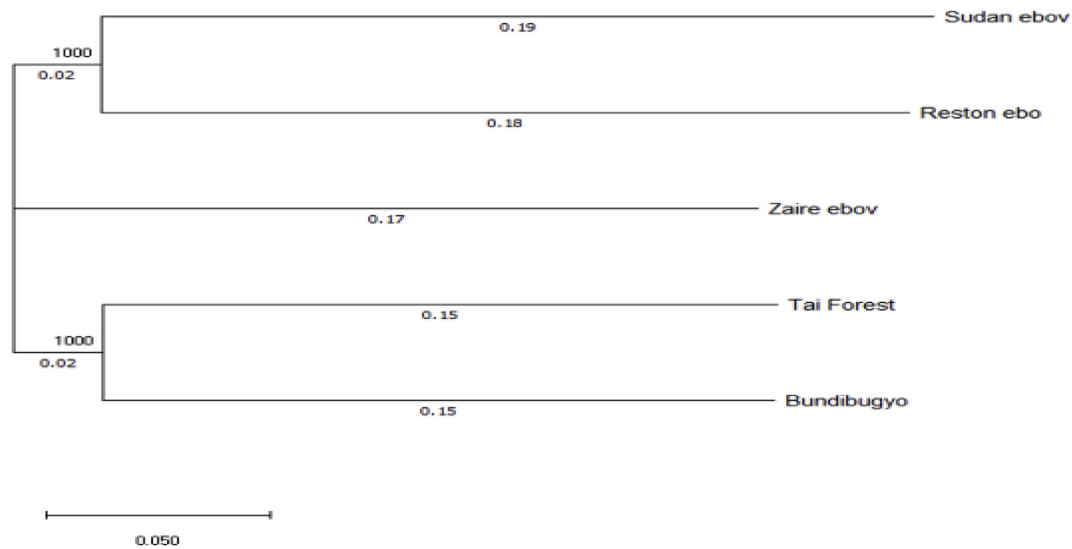
The phylogenetic tree results, from the comparative analysis of the five different strains of Ebola virus, is depicted in Figs. 4–7. The results showed that *Tai forest Ebola virus* and *Bundibugyo Ebola virus* are closely related. The results also showed that the *Sudan and Reston Ebola viruses* are closely related. The *Zaire Ebola virus* stood out from all.

#### 3.2. Results of the evaluation of the phylogenetic tree

As an extended work, the phylogenetic tree produced from the bioinformatics analysis was evaluated. The results of the evaluation of the phylogenetic tree can be found as shown in Figs. 8–13. *Sudan Ebola virus* has a phylogenetic tree evaluation value of 0.19, the *Reston Ebola virus* value is 0.18, *Zaire Ebolarvirus* has 0.17, *Tai forest Ebola virus* has 0.15, and *Bundibugyo Ebola virus* has 0.15.



Fig. 7. Phylogenetic tree, depicting the relationships among the five Ebola virus strains, viewed using iTOL software.



**Fig. 8.** Rectangular cladogram view of the phylogenetic tree evaluation results. This depicts the relationships among the five Ebola virus strains. It was generated via MEGA X software [159]. It was viewed in the newick version using Treeviewer [160]. The bootstrap value used during the evaluation was 1000. This is evident at the nodes of Fig. 8. This shows that the evaluation is good. The smaller numbers on the tree are phylogenetic evaluation estimates.

#### 4. Discussion

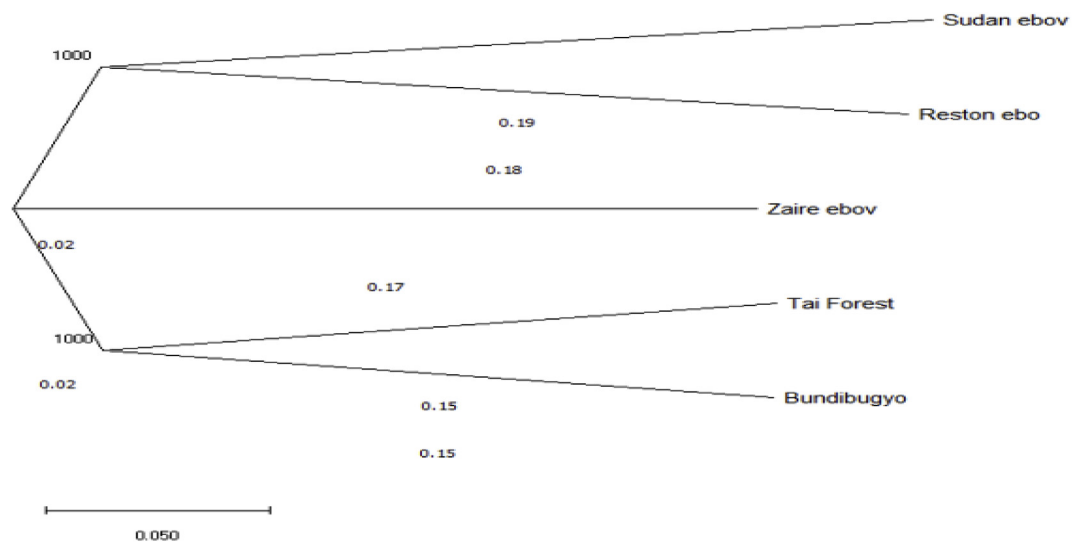
##### 4.1. Discussion on the results of the systematic review

The results obtained from the systematic literature review, has helped identify literature that has been tailored towards controlling the spate of Ebola. The control of EVD can be achieved through useful interventions such as drug targets, vaccines, and immunotherapeutic approaches, amongst others.

Out of the initial 237,498 publications identified, only **104** articles met the inclusion criteria. It is interesting to note that drugs and therapeutics-related Ebola control research, with a total of 41, had the highest number of publications [8–112]. The second control group, classified as other experiments, ranked second, with a total of 28 research publications [8–112]. Vaccine-related Ebola control research ranked third, with a total of 23 research publications [8–112]. This was followed by modeling and simulation-related EVD control research, with a total of 12 research publications [8–112].

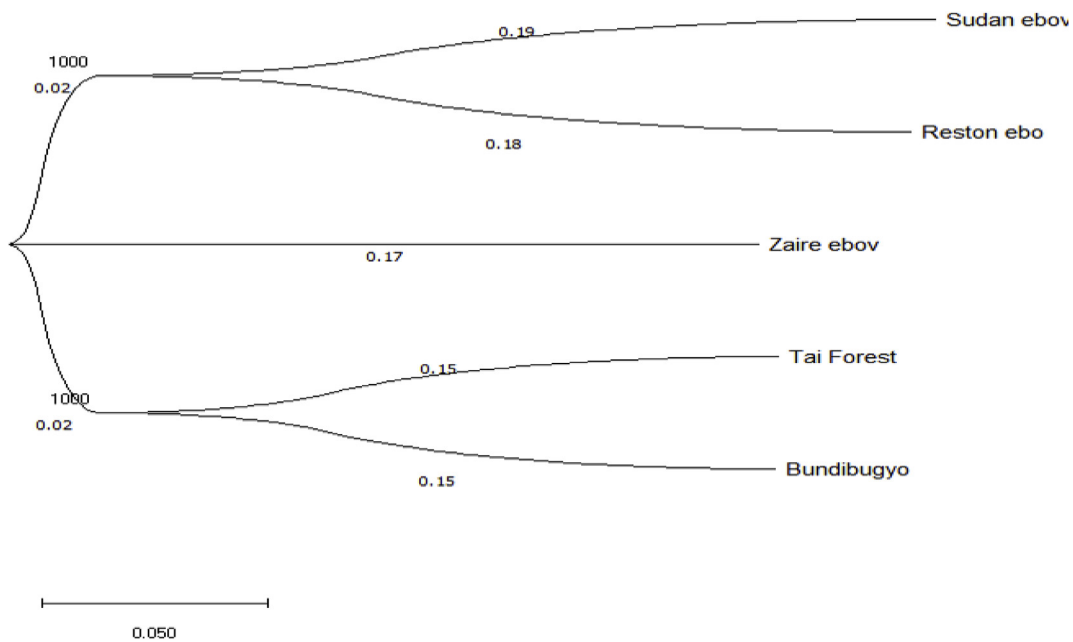
These results reveal that, from 2008 to 2018, there are very few modeling and simulation research articles relating to Ebola control. Modeling and simulation research exhibited the least number of publications. It is evident that no modeling and simulation research studies have been specifically conducted with regard to the impact of multi-level interventions on EVD control, reduction, and possible eradication. Such multi-level interventions include the joint application of multi-target vaccines, multi-target drugs, and multi-target therapeutics toward EVD reduction and possible eradication, on selected communities. This calls for further research. It is also evident from the results of this study, that very few modeling and simulation studies have been conducted on the control of EVD in the last 10 years. Thus, there is need for more modeling and simulation-related Ebola control research. There is a need for more studies on modeling and simulation to assess or estimate the impact of drugs targets, hybrid drugs, and hybrid therapeutic formulations on Ebola incidence, reduction, and possible eradication.

In the last decade, research on drugs or drug targets and therapeutics have contributed immensely to control measures against EVD.



**Fig. 9.** Straight tree cladogram view of the phylogenetic tree evaluation results. This depicts the relationships among the five Ebola virus strains. It was generated by the MEGA X software [159]. It was viewed in the newick version using Treeviewer [160]. The bootstrap value used during the evaluation was 1000. This is evident at the nodes of Fig. 9. This shows that the evaluation is good. The smaller numbers on the tree are phylogenetic evaluation estimates.





**Fig. 10.** Curved cladogram (Bow-shaped view) phylogenetic tree evaluation results. This depicts the relationships among the five Ebola virus strains. It was generated by the MEGA X software [159]. It was viewed in the newick version using Treeviewer [160]. The bootstrap value utilized during the evaluation was 1000. This is evident at the nodes of Fig. 10. This provides evidence that the evaluation is satisfactory. The smaller numbers on the tree are phylogenetic evaluation estimates.

Vaccine-related research have also contributed meaningfully to control efforts against EVD. Modeling and simulation will likely be useful to forecast future outbreaks of EVD. It will also be useful to predict the level of future interventions needed to drastically reduce or avert future EVD outbreaks.

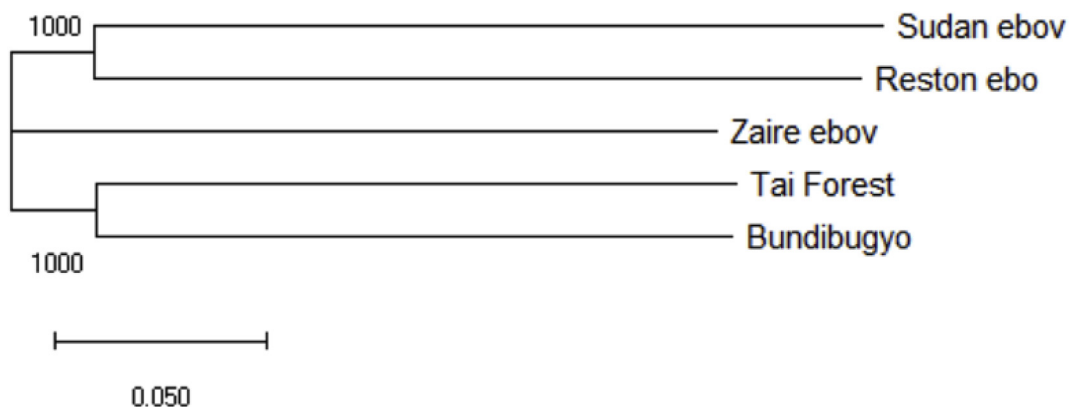
More research is needed to translate Ebola drug-targets, multi-drug targets, and therapeutic targets, into effective EVD drugs. Moreover, modeling and simulation studies can be used for modeling Ebola drug trial designs, therapeutic trial designs, and multi-target vaccine trial designs. Ebola drugs and therapeutic trials are essential for safety and efficacy. Modeling and simulation studies can complement this process.

Another interesting finding revealed that scientists were actively engaged in EVD-control research whenever there were incidences of Ebola Virus Disease. However, whenever there was a lack of EVD incidence, scientists engaged to a lesser extent in EVD-control research. This is evident in Fig. 3. Frequency of EVD-control scientific publications soared higher in years 2015 and 2016, when the 2014–2015 Ebola pandemic ravaged some West African countries. The pandemic caused some panic, and mortality rates were relatively high. It should be noted

that research in scientific publications published in 2015 and 2016 were initially conducted in year 2014 and 2015 respectively (this was a period of Ebola pandemic and high mortality rate). The same applies to scientific publications published in 2017. There had been incidences of EVD in DR Congo in 2016 and most recently in 2017. The incidence accounted for the deaths of many people. This confirms the results obtained in Fig. 3.

#### 4.2. Discussion on the results of the bioinformatics analysis - phylogenetic trees

Fig. 4, depicts the relationship between the five EVD virus strains. It is evident that *Zaire Ebola virus* was distinct from the other four Ebola virus strains. *Zaire Ebola virus* is the most virulent of all Ebola viruses [122]. Consistent with other phylogenetic studies [48,121], our results showed that *Bundibugyo* and *Tai forest Ebola virus* are closely related [See Fig. S5 in [48]; See Fig. 3 in [121]]. *Reston* and *Sudan Ebola virus* are also closely related from the results shown in the phylogenetic tree information. This is similar to the results obtained in prior work. An

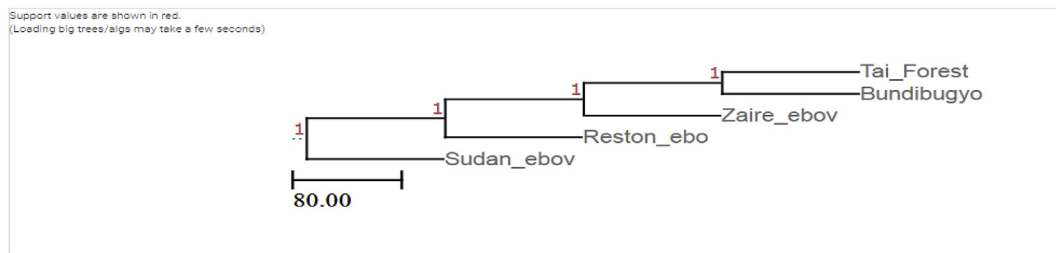


**Fig. 11.** Rectangular Cladogram (Fourth view) of phylogenetic tree evaluation results. This depicts the relationships among the five Ebola virus strains. It was generated by the MEGA X software [159]. It was viewed in the newick version using Treeviewer [160]. The bootstrap value used during the evaluation was 1000. This is evident at the nodes of Fig. 11. This provides evidence that the evaluation is satisfactory. The smaller numbers on the tree are phylogenetic evaluation estimates.

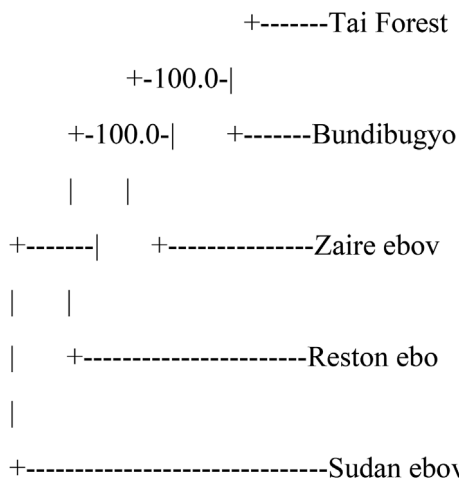


**Table 2**  
Five complete genome of Ebola virus.

Genome	Accession	Source information	Host	Genome Size	Proteins	Neighbors	Date Created	Date Updated
<i>Bundibugyo Ebola virus</i>	NC_014373	isolate: Bundibugyo virus/H.sapiens-tc/UGA/2007/Butalya-811250	Vertebrates, human	18940 nt	9	6	08/09/2010	04/24/2018
<i>Reston Ebola virus</i>	NC_004161	isolate:Reston virus/M.fascicularis-tc/USA/1989/Philippines89-Pennsylvania	Vertebrates, human	18891 nt	9	17	09/04/2002	02/16/2018
<i>Sudan Ebola virus</i>	NC_006432	isolate:Sudan virus/H.sapiens-tc/UGA/2000/Gulu-808892	Vertebrates, human	18875 nt	9	14	11/15/2004	02/16/2018
<i>Tai Forest Ebola virus</i>	NC_014372	isolate:Tai Forest virus/H.sapiens-tc/CIV/1994/Pauleoula-CI	Vertebrates, human	18935 nt	9	1	08/05/2010	04/20/2016
<i>Zaire Ebola virus</i>	NC_002549	isolate:Ebola virus/H.sapiens-tc/COD/1976/Yambuku-Mayinga	Vertebrates, human	18959 nt	9	1382	02/10/1999	02/16/2018



**Fig. 12.** The output from Phylip phylogenetic tree evaluation [158]. It was viewed by using the online Tree viewer [160].



**Fig. 13.** Consensus Tree. The numbers on the branches indicate the number of times the partition of the species into the two sets which are separated by that branch occurred among the trees, out of 100.00 trees.

**Table 3**  
Classification of synthesized Ebola control studies into Potential Control/intervention groups.

Ebola Control Group	Vaccines	Modeling and Simulations	Drugs and Therapeutics	Other Experimental studies
Number of studies from 2008 to 2018	23	12	41	28

**Table 4**  
Showing the frequency of publications of EVD control research between 2008 and 2018.

Year	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018
Number of Publications	1	1	1	1	0	1	2	21	41	32	3

important insight for EVD control is that similar methods of control can be applied to curb the spread of *Reston and Sudan Ebola viruses*, because these viruses are closely related. Similar approaches can also be adopted and applied to *Bundibugyo and Tai forest Ebola viruses*, since these two Ebola viruses are closely related. It may be possible to adopt similar EVD control measures, at the molecular level, against other Ebola virus strains that are closely related. Furthermore, this can facilitate the development and production of joint, multi-protective, multi-treatment drugs and vaccines against Ebola virus strains.

Zaire is the most virulent of the five Ebola viruses [122]. More intensive research efforts should be directed at preventing the transmission of *Zaire Ebola virus*, especially to assist in preventing a major Ebola pandemic from occurring. EVD-control vaccination-related research, drug target-related, and therapeutic-related research, should be encouraged and funded. One of the possible control insights from the phylogenetic analysis result is that molecular and biological experiments on Ebola virus strains that are closely related, should be funded towards discovering hybrid novel control measures against EVD transmissions. Vaccine development and trials have shown great promise in the fight against *Zaire Ebola virus* [10,18,20,23,34,47,96,106]. More research is needed to develop an effective, efficient and safe vaccine against *Zaire Ebola virus*. This current study is potentially significant in the fight against Ebola Virus Disease (EVD).

4.3. Discussion on the evaluation of the phylogenetic tree

Figs. 8–13, depict the results of the evaluations of the phylogenetic tree. It is evident that the outputs from the MEGA X phylogenetic tree evaluation, and the Phylip phylogenetic tree evaluations, yielded the same results. It is evident that these results can spur more research toward the control of EVD. The values obtained from the evaluation of the phylogenetic trees reveal that some of the virus strains are very

closely related. Such results can provide a platform for further research into the evolutionary analysis of the five different Ebola viruses. Such analysis may produce novel results. The results may then be useful to ultimately eradicate EVD. Furthermore, the results obtained from the evaluation of the phylogenetic tree, can help provide insight into the origins and evolution of these Ebola viruses. The evaluation can also provide insight into the possible structural and genetic mutations that each Ebola virus may undergo. Finally, the structural and functional properties of the genetic make-up of the genes of each Ebola virus can be inferred. Knowledge from such inference can be channeled toward the control of EVD.

## 5. Recommendations

Proper partnership with relevant health organizations can be assistive in future surveillance efforts. This will help in the fight against EVD. Prompt response to an EVD outbreak, will help in reducing the transmission of the disease. Isolation and treatment of EVD infected individuals, will help in curtailing the spread of the disease. Investment and adequate funding for biological computation, bioinformatics and molecular research on EVD, will assist scientists to develop novel disease treatments. Regulation on the consumption of bush meat can also help to reduce the transmission of EVD. Implementation strategies, computational models and computational systems that can send early signals and predict possible future outbreaks of EVD should be implemented. Educating and sensitizing the public, both rural and urban dwellers, concerning the prevention of Ebola Virus Disease would be a substantive progress. Effective border control and health screening at various ports of entry for all African countries, as well as other nations, should be implemented. All of these recommendations, if fully implemented, can play a major role in the effective control of EVD.

## 6. Conclusion

This study has provided generalized and in-silico insight for the control of EVD from the studies identified according to inclusion criteria. To the best of our knowledge, the investigation has extensively reviewed the Ebola control-related literature. Insight for Ebola control was gained from the reviewed studies. Insight for EVD control was also obtained from the bioinformatics analyses performed on five different strains of Ebola virus. Evaluation of the phylogenetic tree provided further insight for EVD control. We recommend that scientists should continue these efforts toward EVD control. Investigators should consistently engage in active research toward EVD control and possible eradication.

## Conflicts of interest

The authors declare that there are no competing interests.

## Author contribution

Olugbenga Oluwagbemi conceived of the research and designed the overall study. Awe Olaitan and Olugbenga Oluwagbemi performed the bioinformatics analysis; Olugbenga Oluwagbemi wrote the manuscript. Olugbenga Oluwagbemi conducted the systematic review and rewrote the revised version of the manuscript. All authors read and approved the final manuscript.

## Ethical statement

The names of authors that contributed to the paper have been included. We have conducted plagiarism check on the manuscript. We used the turnitin plagiarism checker for this purpose. The result was OK. The paper has not been submitted to any other journal for consideration. All references have been duly cited.

## Acknowledgement

This work is based on the research supported partly/wholly by the National Research Foundation (NRF) of South Africa, (With Grant Number: UID: 111988) of the DST/NRF Innovation Postdoctoral Research Grant Award, awarded to Olugbenga Oluwagbemi.

The grant holder acknowledges that opinions, findings, and conclusions or recommendations expressed in this publication of this NRF supported research, is that of the authors and that the NRF accepts no liability whatsoever in this regard.

We thank Dr. Ifeyinwa Okolo, of Stellenbosch University, South Africa, for taking time to correct the English language constructs of the manuscript. We also thank Olaoluwa Durobello, of the University of Nottingham in Malaysia, for taking time to correct the English language constructs of the manuscript.

## Appendix A. Supplementary data

Supplementary data related to this article can be found at <https://doi.org/10.1016/j.imu.2018.07.004>.

## References

- [1] Raka L, Guardo M. Ebola in West Africa. open access Macedonian. *J Med Sci* 2015;3(1). 1–174-175.
- [2] WHO 2017; <http://www.who.int/csr/don/13-may-2017-Ebola-drc/en/>; Access date: 13th May 2017.
- [3] Holmes EC, Dudas Gytis, Rambaut Andrew, Andersen Kristian G. The evolution of Ebola virus: insights from the 2013–2016 epidemic. *Nature* 2016;538:193–200. (13 October 2016) doi:10.1038/nature19790.
- [4] Omilabu SA, Salu OB, Oke BO, James AB. The West African Ebola virus disease epidemic 2014–2015: a commissioned review. *Niger Postgrad Med J* 2016;23(2):49–56.
- [5] Martínez MJ, Salim Abdulbaset M, Hurtado Juan C, Kilgore Paul E. Ebola virus infection: overview and update on prevention and treatment. *Infect Dis Ther* 2015;4(4):365–90.
- [6] Mendoza EJ, Qiu Xiangguo, Kobinger Gary P. Progression of Ebola therapeutics during the 2014–2015 outbreak. *Trends Mol Med* 2016;22(2):164–73.
- [7] WHO 2018; <http://www.who.int/csr/don/10-may-2018-Ebola-drc/en/>; Access date: 13th May, 2018.
- [8] Borisevich IV, Chemikova NK, Markov VI, Krasnianskiy VP, Borisevich SV, Rozhdstvenskiy EV. An experience in the clinical use of specific immunoglobulin from horse blood serum for prophylaxis of Ebola haemorrhagic fever. *Vopr Virusol* 2017;62(1):25–9. 2017.
- [9] Raftery P, Condell O, Wasunna C, Kpaka J, Zwizwai R, Nuha M, et al. Establishing Ebola Virus Disease (EVD) diagnostics using GeneXpert technology at a mobile laboratory in Liberia: impact on outbreak response, case management and laboratory systems strengthening. *PLoS Neglected Trop Dis* 2018;12(1):e0006135 <https://doi.org/10.1371/journal.pntd.0006135>.
- [10] Kennedy SB, Bolay F, Kieh M, Grandits G, Badio M, Ballou R, Eckes R, Feinberg M, Follmann D, Grund B, Gupta S, Hensley L, Higgs E, Janosko K, Johnson M, Kateh F, Logue J, Marchand J, Monath T, Nason M, Nyenswah T, Roman F, Stavale E, Wolfson J, Neaton JD, Lane HC. PREVAIL I study group. *N Engl J Med* 2017;377(15):1438–47. <https://doi.org/10.1056/NEJMoa1614067>.
- [11] Agnandji ST, Fernandes JF, Bache EB, Obiang Mba RM, Brosnahan JS, Kabwende L, Pitzinger P, Staarink P, Massinga-Loembe M, Krähling V, Biedenkopf N, Fehling SK, Strecker T, Clark DJ, Staines HM, Hooper JW, Silvera P, Moorthy V, Kiény MP, Adegnikaa AA, Grobusch MP, Becker S, Ramharther M, Mordmüller B, Lell B, VEBCON Consortium, Krishna S, Krensner PG. Safety and immunogenicity of rVSVΔG-ZEBOV-GP Ebola vaccine in adults and children in Lambaréné, Gabon: a phase I randomised trial. *PLoS Med*. 2017;14(10):e1002402 <https://doi.org/10.1371/journal.pmed.1002402>. 2017 Oct 6.
- [12] Mire CE, Geisbert TW. Neutralizing the threat: Pan-Ebolavirus antibodies close the loop. *Trends Mol Med* 2017;23(8):669–71. <https://doi.org/10.1016/j.molmed.2017.06.008>.
- [13] ElSherif MS, Brown C, MacKinnon-Cameron D, Li L, Racine T, Alimonti J, Rudge TL, Sabourin C, Silvera P, Hooper JW, Kwilas SA, Kilgore N, Badorrek C, Ramsey WJ, Heppner DG, Kemp T, Monath TP, Nowak T, McNeil SA, Langley JM, Halperin SA. Canadian Immunization Research Network. Assessing the safety and immunogenicity of recombinant vesicular stomatitis virus Ebola vaccine in healthy adults: a randomized clinical trial. *Can Med Assoc J* 2017;189(24):E819–27. <https://doi.org/10.1503/cmaj.170074>.
- [14] Heppner Jr. DG, Kemp TL, Martin BK, Ramsey WJ, Nichols R, Dasen EJ, Link CJ, Das R, Xu ZJ, Sheldon EA, Nowak TA, Monath TP. V920-004 study team. Safety and immunogenicity of the rVSVΔG-ZEBOV-GP Ebola virus vaccine candidate in healthy adults: a phase 1b randomised, multicentre, double-blind, placebo-controlled, dose-response study. *Lancet Infect Dis* 2017;17(8):854–66. [https://doi.org/10.1016/S1473-3099\(17\)30313-4](https://doi.org/10.1016/S1473-3099(17)30313-4). 2017 Aug.
- [15] Halperin SA, Arribas JR, Rupp R, Andrews CP, Chu L, Das R, Simon JK, Onorato

- MT, Liu K, Martin J, Helmond FA. V920-012 Study Team. Six-month safety data of recombinant vesicular stomatitis virus-zaire Ebola virus envelope glycoprotein vaccine in a phase 3 double-blind, placebo-controlled randomized study in healthy adults. *J Infect Dis* 2017;215(12):1789–98. 15 June 2017, Pages.
- [16] Gallandat K, Lantagne D. Selection of a Biosafety Level 1 (BSL-1) surrogate to evaluate surface disinfection efficacy in Ebola outbreaks: comparison of four bacteriophages. *PLoS One* 2017;12(5):e0177943 <https://doi.org/10.1371/journal.pone.0177943>. eCollection 2017. 2017 May 22.
- [17] Zhang Y, Gong Y, Wang C, Liu W, Wang Z, Xia Z, Bu Z, Lu H, Sun Y, Zhang X, Cao Y, Yang F, Su H, Hu Y, Deng Y, Zhou B, Zhao Z, Fu Y, Kargbo D, Dfafe F, Kargbo B, Kanu A, Liu L, Qian J, Guo Z. Rapid deployment of a mobile biosafety level-3 laboratory in Sierra Leone during the 2014 Ebola virus epidemic. *PLoS Neglected Trop Dis* 2017;11(5). <https://doi.org/10.1371/journal.pntd.0005622>. eCollection 2017 May. e0005622.
- [18] Medaglini D, Siegrist CA. Immunomonitoring of human responses to the rVSV-ZEBOV Ebola vaccine. *Curr Opin Virol* 2017;23:88–94. <https://doi.org/10.1016/j.coviro.2017.03.008>.
- [19] Dudas G, Carvalho LM, Bedford T, Tatem AJ, Baele G, Faria NR, Park DJ, Ladner JT, Arias A, Asogun D, Bielejec F, Caddy SL, Cotten M, D'Ambrozio J, Dellicour S, Di Caro A, DiClaro JW, Duraffour S, Elmoro MJ, Fakoli LS, Faye O, Gilbert ML, Gevao SM, Gire S, Gladden-Young A, Gnirke A, Goba A, Grant DS, Haagmans BL, Hiscox JA, Jah U, Kugelman JR, Liu D, Lu J, Malboeuf CM, Mate S, Matthews DA, Matranga CB, Meredith LW, Qu J, Quick J, Pas SD, Phan MVT, Pollakis G, Reusken CB, Sanchez-Lockhart M, Schaffner SF, Schieffelin JS, Sealfon RS, Simon-Loriere E, Smits SL, Stoecker K, Thorne L, Tobin EA, Vandi MA, Watson SJ, West K, Whitmer S, Wiley MR, Winnicki SM, Wohl S, Wölfel R, Yozviak NL, Andersen KG, Blyden SO, Bolay F, Carroll MW, Dahn B, Diallo B, Formenty P, Fraser C, Gao GF, Garry RF, Goodfellow I, Günther S, Happi CT, Holmes EC, Kargbo B, Keita S, Kellam P, Koopmans MPG, Kuhn JH, Loman NJ, Magassouba N, Naidoo D, Nichol ST, Nyenswah T, Palacios G, Pybus OG, Sabeti PC, Sall A, Ströher U, Wurie I, Suchard MA, Lemey P, Rambaut A. Virus genomes reveal factors that spread and sustained the Ebola epidemic. *Nature* 2017;544(7650):309–15. <https://doi.org/10.1038/nature22040>. 2017 Apr 20.
- [20] Huttner A, Combesure C, Grillet S, Haks MC, Quinten E, Modoux C, Agnandji ST, Brossnahan J, Dayer JA, Harandi AM, Kaiser L, Medaglini D, Monath T, VEBCON, VSV-EBOVAC Consortia Roux-Lombard P, Kremsner PG, Ottenhoff TH, Siegrist CA. A dose-dependent plasma signature of the safety and immunogenicity of the rVSV-Ebola vaccine in Europe and Africa. *Sci Transl Med* 2017;9(385). <https://doi.org/10.1126/scitranslmed.aaj1701>. pii: eaaj1701.
- [21] Funk S, Ciglenecki I, Tiffany A, Gignoux E, Camacho A, Eggo RM, Kucharski AJ, Edmunds WJ, Bolongee J, Azuma P, Clement P, Alpha TS, Sterk E, Telfer B, Engel G, Parker LA, Suzuki M, Heijenberg N, Reeder B. The impact of control strategies and behavioural changes on the elimination of Ebola from Lofa County, Liberia. *Philos Trans R Soc Lond B Biol Sci* 2017;372(1721). <https://doi.org/10.1098/rstb.2016.0302>. pii: 20160302.
- [22] Gallandat K, Wolfe MK, Lantagne D. Surface cleaning and disinfection: efficacy assessment of four chlorine types using *Escherichia coli* and the Ebola surrogate Phi6. *Environ Sci Technol* 2017;51(8):4624–31. <https://doi.org/10.1021/acs.est.6b06014>. 2017 Apr 18.
- [23] Winslow RL, Milligan ID, Voysey M, Luhn K, Shukarev G, Douoguih M, Snape MD. Immune responses to novel adenovirus type 26 and modified vaccinia virus ankara-vectored Ebola vaccines at 1 year. *J Am Med Assoc* 2017;317(10):1075–7. <https://doi.org/10.1001/jama.2016.20644>.
- [24] Wolfe MK, Gallandat K, Daniels K, Desmarais AM, Scheinman P, Lantagne D. Handwashing and Ebola virus disease outbreaks: a randomized comparison of soap, hand sanitizer, and 0.05% chlorine solutions on the inactivation and removal of model organisms Phi6 and *E. coli* from hands and persistence in rinse water. *PLoS One* 2017;12(2):e0172734 <https://doi.org/10.1371/journal.pone.0172734>.
- [25] Konde MK, Baker DP, Traore FA, Sow MS, Camara A, Barry AA, Mara D, Barry A, Cone M, Kaba I, Richard AA, Beavogui AH, Günther S, European Mobile Laboratory Consortium. Pintilie M, Fish EN. Interferon  $\beta$ -1a for the treatment of Ebola virus disease: a historically controlled, single-arm proof-of-concept trial. *PLoS One* 2017;12(2). <https://doi.org/10.1371/journal.pone.0169255>. eCollection 2017. e0169255.
- [26] Li H, Yu F, Xia S, Yu Y, Wang Q, Lv M, Wang Y, Jiang S, Lu L. Chemically modified human serum albumin potentially blocks entry of Ebola pseudoviruses and viruslike particles. *Antimicrob Agents Chemother* 2017;61(4). <https://doi.org/10.1128/AAC.021168-16>. pii: e021168-16.
- [27] Dolzhikova IV, Zubkova OV, Tukhvatulin AI, Dzharullaeva AS, Tukhvatulina NM, Shcheplyakov DV, Shmarov MM, Tokarskaya EA, Simakova YV, Egorova DA, Scherbinin DN, Tutukhina IL, Lysenko AA, Kostornoy AV, Gancheva PG, Ozharovskaya TA, Belugin BV, Kolobukhina LV, Pantyukhov VB, Syromyatnikova SI, Shatokhina IV, Sizikova TV, Rumyantseva IG, Andrus AF, Boyarskaya NV, Voytyuk AN, Babira VF, Volchikhina SV, Kutaev DA, Bel'skikh AN, Zhdanov KV, Zakharenko SM, Borisevich SV, Logunov DY, Naroditsky BS, Gintsburg AL. Safety and immunogenicity of GamEvac-Comb1, a heterologous VSV- and Ad5-vectored Ebo vaccine: an open phase I/II trial in healthy adults in Russia. *Hum Vaccines Immunother* 2017;13(3):613–20. <https://doi.org/10.1080/21645515.2016.1238535>. 2017 Mar 4.
- [28] Siegel D, Hui HC, Doerffler E, Clarke MO, Chun K, Zhang L, Neville S, Carra E, Lew W, Ross B, Wang Q, Wolfe L, Jordan R, Soloveva V, Knox J, Perry J, Perron M, Stray KM, Barauskas O, Feng JY, Xu Y, Lee G, Rheingold AL, Ray AS, Bannister R, Strickley R, Swaminathan S, Lee WA, Bavari S, Cihlar T, Lo MK, Warren TK, Mackman RL. Discovery and synthesis of a phosphoramidate prodrug of a pyrrolo [2,1-f][triazin-4-amino] adenine C-Nucleoside (GS-5734) for the treatment of Ebola and emerging viruses. *J Med Chem* 2017;60(5):1648–61. <https://doi.org/10.1021/acs.jmedchem.6b01594>.
- [29] Ueda MT, Kurosaki Y, Izumi T, Nakano Y, Oloniyi OK, Yasuda J, Koyanagi Y, Sato K, Nakagawa S. Functional mutations in spike glycoprotein of Zaire Ebola virus associated with an increase in infection efficiency. *Gene Cell* 2017;22(2):148–59. <https://doi.org/10.1111/gtc.12463>. 2017 Feb.
- [30] Liang J, Sagum CA, Bedford MT, Sidhu SS, Sudol M, Han Z, Harty RN. Chaperone-Mediated autophagy protein BAG3 negatively regulates Ebola and marburg VP40-mediated egress. *PLoS Pathog* 2017;13(7):e1006519 <https://doi.org/10.1371/journal.ppat.1006519>.
- [31] Dörnemann J, Burzio C, Ronsse A, Sprecher A, De Clerck H, Van Herp M, Kolić MC, Yosifiva V, Caluwaerts S, McElroy AK, Antierens A. First newborn baby to receive experimental therapies survives Ebola virus disease. *J Infect Dis* 2017;215(2):171–4. <https://doi.org/10.1093/infdis/jiw493>. PMID:28073857. 2017 Jan 15.
- [32] Li JX, Hou LH, Meng FY, Wu SP, Hu YM, Liang Q, Chu K, Zhang Z, Xu JJ, Tang R, Wang WJ, Liu P, Hu JL, Luo L, Jiang R, Zhu FC, Chen W. Immunity duration of a recombinant adenovirus type-5 vector-based Ebola vaccine and a homologous prime-boost immunisation in healthy adults in China: final report of a randomised, double-blind, placebo-controlled, phase 1 trial. *Lancet Glob Health* 2017;5(3):e324–34. [https://doi.org/10.1016/S2214-109X\(16\)30367-9](https://doi.org/10.1016/S2214-109X(16)30367-9). 2017 Mar.
- [33] Henaó-Restrepo AM, Camacho A, Longini IM, Watson CH, Edmunds WJ, Egger M, Carroll MW, Dean NE, Diatta I, Doumbia M, Draguez B, Duraffour S, Enwere G, Grais R, Gunther S, Gsell PS, Hossmann S, Watle SV, Kondé MK, Kéita S, Kone S, Kuisma E, Levine MM, Mandal S, Maugé T, Norheim G, Riveros X, Soumah A, Trelle S, Vicari AS, Rottingen JA, Kiemy MP. Efficacy and effectiveness of an rVSV-vectored vaccine in preventing Ebola virus disease: final results from the Guinea ring vaccination, open-label, cluster-randomised trial (Ebola Ça Suffit!). *Lancet*. 2017 Feb 4;389(10068):504. [https://doi.org/10.1016/S0140-6736\(16\)03036-9](https://doi.org/10.1016/S0140-6736(16)03036-9). 2017 Feb 4.
- [34] Shukarev G, Callendret B, Luhn K, Douoguih M. EBOVAC1 consortium. A two-dose heterologous prime-boost vaccine regimen eliciting sustained immune responses to Ebola Zaire could support a preventive strategy for future outbreaks. *Hum Vaccines Immunother* 2017;13(2):266–70. <https://doi.org/10.1080/21645515.2017.1264755>. 2017 Feb.
- [35] Cong Y, Dyaal J, Hart BJ, DeWald LE, Johnson JC, Postnikova E, et al. Evaluation of the activity of lamivudine and zidovudine against Ebola virus. *PLoS One* 2016;11(11):e0166318 <https://doi.org/10.1371/journal.pone.0166318>.
- [36] Merler S, Ajelli M, Fumanelli L, Parlamento S, Pastore y Piontti A, Dean NE, et al. Containing Ebola at the source with ring vaccination. *PLoS Neglected Trop. Dis.* 2016;10(11):e0005093 <https://doi.org/10.1371/journal.pntd.0005093>.
- [37] Dowall SD, Bewley K, Watson RJ, Vasan SS, Ghosh C, Konai MM, Gausdal G, Lorens JB, Long J, Barclay W, Garcia-Dorival I, Hiscox J, Bosworth A, Taylor I, Easterbrook L, Pitman J, Summers S, Chan-Pensley J, Funnell S, Vipond J, Charlton S, Haldar J, Hewson R, Carroll MW. Antiviral screening of multiple compounds against Ebola virus. *Viruses* 2016;8(11). 2016 Oct 27; pii: E277. PMID:27801778.
- [38] Bai CQ, Mu JS, Kargbo D, Song YB, Niu WK, Nie WM, Kanu A, Liu WW, Wang YP, Dfafe F, Yan T, Hu Y, Deng YQ, Lu HJ, Yang F, Zhang XG, Sun Y, Cao YX, Su HX, Sun Y, Liu WS, Wang CY, Qian J, Liu L, Wang H, Tong YG, Liu ZY, Chen YS, Wang HQ, Kargbo B, Gao GF, Jiang JF. Clinical and virological characteristics of Ebola virus disease patients treated with Favipiravir (T-705)-Sierra Leone, 2014. *Clin Infect Dis* 2016;63(10):1288–94. 2016 Nov 15.
- [39] Welch SR, Guerrero LW, Chakrabarti AK, McMullan LK, Flint M, Bluemling GR, Painter GR, Nichol ST, Spiropoulou CF, Albarrão CG. Lassa and Ebola virus inhibitors identified using minigenome and recombinant virus reporter systems. *Antivir Res* 2016;136:9–18. <https://doi.org/10.1016/j.antiviral.2016.10.007>.
- [40] Li ZJ, Tu WX, Wang XC, Shi GQ, Yin ZD, Su HJ, Shen T, Zhang DP, Li JD, Lv S, Cao CL, Xie RQ, Lu HZ, Jiang RM, Cao Z, An ZJ, Li LL, Xu J, Xiong YW, Zhang W, Zhang W, Zhang HW, Chen WS, Ling H, Xu W, Cai J, Luo HJ, Xing XS, Zheng CJ, Wei Q, Li XX, Li M, Jiang H, Deng LQ, Chen MQ, Huo X, Xu F, Lai XH, Bai XC, Ye LJ, Yao JY, Yin WW, Sun JJ, Xiao L, Liu FQ, Liu XQ, Fan HW, Kou ZQ, Zhou JK, Zhang H, Ni DX, Samba TT, Li Q, Yu HJ, Wang Y, Liang XF. A practical community-based response strategy to interrupt Ebola transmission in Sierra Leone, 2014-2015. *Infect Dis Poverty* 2016;5(1):74. <https://doi.org/10.1186/s40249-016-0167-0>. 2016 Aug 5.
- [41] Olsen ME, Filone CM, Rozelle D, Mire CE, Agans KN, Hensley L, Connor JH. Polyamines and hypusination are required for ebolavirus gene expression and replication. *mBio* 2016;7(4). <https://doi.org/10.1128/mBio.00882-16>. 2016 Jul 26;pii: e00882-16.
- [42] Ahmad MD, Usman M, Khan A, Imran M. Optimal control analysis of Ebola disease with control strategies of quarantine and vaccination. *Infect Dis Poverty* 2016;5(1):72. <https://doi.org/10.1186/s40249-016-0161-6>.
- [43] Cohen NJ, Brown CM, Alvarado-Ramy F, Bair-Brake H, Benenson GA, Chen TH, Demma AJ, Holton NK, Kohl KS, Lee AW, McAdam D, Pesik N, Roohi S, Smith CL, Waterman SH, Cetron MS. Travel and border health measures to prevent the international spread of Ebola. *MMWR* 2016;65(3):57–67. <https://doi.org/10.15585/mmwr.su6503a9>. Suppl. 2016 Jul 8.
- [44] Hageman JC, Hazim C, Wilson K, Malpiedi P, Gupta N, Bennett S, Kolwaite A, Tumpey A, Brinsley-Rainisch K, Christensen B, Gould C, Fisher A, Schaefer M, Hamilton D, Moran K, Delaney L, Dowell C, Bell M, Srinivasan A, Jhuam M, Fagan R, Adrien N, Chea N, Park BJ. Infection prevention and control for Ebola in health care settings - West Africa and United States. *MMWR* 2016;65(3):50–6. <https://doi.org/10.15585/mmwr.su6503a8>. Suppl. 2016 Jul 8.
- [45] Mulangu S, Borchert M, Paweska J, Tshomba A, Afounde A, Kulidri A, Swanepoel R, Muyembe-Tamfum JJ, Van der Stuyft P. High prevalence of IgG antibodies to Ebola virus in the efé pygmy population in the watsa region, democratic republic



- of the Congo. *BMC Infect Dis* 2016;16:263. <https://doi.org/10.1186/s12879-016-1607-y>. 2016 Jun 10.
- [46] Gittings K, Matson KL. Establishing herd immunity against Ebola through vaccination. *Vaccine* 2016;34(24):2644–7. <https://doi.org/10.1016/j.vaccine.2016.04.047>.
- [47] Wong KK, Davey Jr. RT, Hewlett AL, Kraft CS, Mehta AK, Mulligan MJ, Beck A, Dorman W, Kratochvil CJ, Lai L, Palmore TN, Rogers S, Smith PW, Suffredini AF, Wolcott M, Ströher U, Uyeky TM. Use of postexposure prophylaxis after occupational exposure to Zaire Ebola virus. *Clin Infect Dis* 2016;63(3):376–9. <https://doi.org/10.1093/cid/ciw256>. Epub 2016 Apr 26. 2016 Aug 1.
- [48] Dhanda SK, Chaudhary K, Gupta S, Brahmachari SK, Raghava GP. A web-based resource for designing therapeutics against Ebola Virus. *Sci Rep* 2016;26(6):24782. <https://doi.org/10.1038/srep24782>. PMID:27113850. 2016 Apr.
- [49] Taylor R, Kotian P, Warren T, Panchal R, Bavari S, Julander J, Dobo S, Rose A, El-Kattan Y, Taubenheim B, Babu Y, Sheridan WP. BCX4430-A broad-spectrum antiviral adenosine nucleoside analog under development for the treatment of Ebola virus disease. *J Infect Public Health* 2016;9(3):220–6. <https://doi.org/10.1016/j.jiph.2016.04.002>. 2016 May-Jun.
- [50] Muñoz A, Sigwalt D, Illescas BM, Luczkowiak J, Rodríguez-Pérez L, Nierengarten I, Holler M, Remy JS, Buffet K, Vincent SP, Rojo J, Delgado R, Nierengarten JF, Martín N. Synthesis of giant globular multivalent glycofullerenes as potent inhibitors in a model of Ebola virus infection. *Nat Chem* 2016;8(1):50–7.
- [51] Cap AP, Pidcoke HF, Keil SD, Staples HM, Anantpadma M, Carrion Jr. R, Davey RA, Frazer-Abel A, Taylor AL, Gonzales R, Patterson JL, Goodrich RP. Treatment of blood with a pathogen reduction technology using ultraviolet light and riboflavin inactivates Ebola virus in vitro. *Transfusion* 2016;56(1):S6–15. <https://doi.org/10.1111/trf.13393>. PMID:27001363.
- [52] Maduka O, Maleghemi S, Komakech W, Nwaduito I, Green P, Ikpe A, Ywoga D, Onyekwene N. Effective risk communication and contact tracing for Ebola virus disease prevention and control - experiences from Port Harcourt, Nigeria. 2016;135:140–3. <https://doi.org/10.1016/j.puhe.2015.10.037>. 2016 Jun.
- [53] Lokuge K, Caleo G, Greig J, Duncombe J, McWilliam N, Squire J, Lamin M, Veltus E, Wolz A, Kobinger G, de la Vega MA, Gbabei O, Nabieu S, Lamin M, Kremer R, Danis K, Banks E, Glass K. Successful control of Ebola virus disease: analysis of service based data from rural Sierra Leone. *PLoS Neglected Trop Dis* 2016;10(3). <https://doi.org/10.1371/journal.pntd.0004498>. e0004498.
- [54] Yasmin and Nabi. B and T Cell epitope-based peptides predicted from evolutionarily conserved and whole protein sequences of Ebola virus as vaccine targets. *Scand J Immunol* 2016;83(5):321–37. <https://doi.org/10.1111/sji.12425>. PMID:26939891.
- [55] Corti D, Misasi J, Mulangu S, Stanley DA, Kanekiyo M, Wollen S, Ploquin A, Doria-Rose NA, Staupé RP, Bailey M, Shi W, Choe M, Marcus H, Thompson EA, Cagigi A, Silacci C, Fernandez-Rodriguez B, Perez L, Sallusto F, Vanzetta F, Agatic G, Camerini E, Kosalu N, Gordon I, Ledgerwood JE, Mascola JR, Graham BS, Muyembe-Tamfun JJ, Trefry JC, Lanzavecchia A, Sullivan NJ. Protective monotherapy against lethal Ebola virus infection by a potentially neutralizing antibody. *Science* 2016;351(6279):1339–42. <https://doi.org/10.1126/science.aad5224>. 2016 Mar 18.
- [56] Misasi J, Gilman MS, Kanekiyo M, Gui M, Cagigi A, Mulangu S, Corti D, Ledgerwood JE, Lanzavecchia A, Cunningham J, Muyembe-Tamfun JJ, Baxa U, Graham BS, Xiang Y, Sullivan NJ, McLellan JS. Structural and molecular basis for Ebola virus neutralization by protective human antibodies. *Science* 2016;351(6279):1343–6. <https://doi.org/10.1126/science.aad6117>.
- [57] Furuyama W, Marzi A, Nanbo A, Haddock E, Maruyama J, Miyamoto H, Igarashi M, Yoshida R, Noyori O, Feldmann H, Takada A. Discovery of an antibody for pan-Ebola virus therapy. *Sci Rep* 2016;10(6):20514. <https://doi.org/10.1038/srep20514>. 2016 Feb.
- [58] Quick J, Loman NJ, Duraffour S, Simpson JT, Severi E, Cowley L, Bore JA, Koundouno R, Dudas G, Mikhail A, Ouedraogo N, Afrough B, Bah A, Baum JH, Doerrbecker J, Enkirch T, Dorival IGG, Heltzel N, Hinzmann J, Holm T, Kafetzopoulou LE, Koropogui M, Kosgey A, Kuisma E, Logue CH, Mazzarelli A, Meisel S, Mertens M, Michel J, Ngabo D, Nitzsche K, Pallash E, Patrono LV, Portmann J, Repits JG, Rickett NY, Sachse A, Singethan K, Vitoriano I, Yemanberhan RL, Zekeng EG, Trina R, Bello A, Sall AA, Faye O, Faye O, Magassouba N, Williams CV, Amburgey V, Winona L, Davis E, Gerlach J, Washington F, Monteil V, Jourdain M, Bererd M, Camara A, Somlare H, Camara A, Gerard M, Bado G, Baillet B, Delaune D, Nebie KY, Diarra A, Savane Y, Pallawo RB, Gutierrez GJ, Milhano N, Roger I, Williams CJ, Yattara F, Lewandowski K, Taylor J, Rachwal P, Turner D, Pollakis G, Hiscox JA, Matthews DA, O'Shea MK, Johnston AM, Wilson D, Hutley E, Smit E, Di Caro A, Woelfel R, Stoecker K, Fleischmann E, Gabriel M, Weller SA, Koivogui L, Diallo B, Keita S, Rambaut A, Formenty P, Gunther S, Carroll MW. Real-time, portable genome sequencing for Ebola surveillance. *Nature* 2016;530(7589):228–32. <https://doi.org/10.1038/nature16996>.
- [59] Zhang X, Ao Z, Bello A, Ran X, Liu S, Wagle J, Kobinger G, Yao X. Characterization of the inhibitory effect of an extract of *Prunella vulgaris* on Ebola virus glycoprotein (GP)-mediated virus entry and infection. *Antivir Res* 2016;127:20–31. <https://doi.org/10.1016/j.antiviral.2016.01.001>. 2016 Mar.
- [60] Wang Y, Cui R, Li G, Gao Q, Yuan S, Altmeyer R, Zou G. Teicoplanin inhibits Ebola pseudovirus infection in cell culture. *Antivir Res* 2015;125:1–7. <https://doi.org/10.1016/j.antiviral.2015.11.003>. 2016 Jan.
- [61] Tapia MD, Sow SO, Lyke KE, Haidara FC, Diallo F, Doumbia M, Traore A, Coulibaly F, Kodio M, Onwuchekwa U, Szein MB, Wahid R, Campbell JD, Kiemy MP, Moorthy V, Imoukhuede EB, Rampling T, Roman F, De Ryck I, Bellamy AR, Dally L, Mbaya OT, Ploquin A, Zhou Y, Stanley DA, Bailer R, Koup RA, Roederer M, Ledgerwood J, Hill AVS, Ballou WR, Sullivan N, Graham B, Levine MM. Use of Chad3-EBO-Z Ebola virus vaccine in Malian and US adults, and boosting of Malian adults with MVA-BN-Filo: a phase 1, single-blind, randomised trial, a phase 1b, open-label and double-blind, dose-escalation trial, and a nested, randomised, double-blind, placebo-controlled trial. *Lancet Infect Dis* 2016;16(1):31–42. [https://doi.org/10.1016/S1473-3099\(15\)00362-X](https://doi.org/10.1016/S1473-3099(15)00362-X). 2016 Jan.
- [62] Kucharski AJ, Eggo RM, Watson CH, Camacho A, Funk S, Edmunds WJ. Effectiveness of ring vaccination as control strategy for Ebola virus disease. *Emerg Infect Dis* 2016 Jan;22(1):105–8. <https://doi.org/10.3201/eid2201.151410>. PMID:26691346.
- [63] McMullan LK, Flint M, Dyall J, Albariño C, Olinger GG, Foster S, Sethna P, Hensley LE, Nichol ST, Lanier ER, Spiropoulos CF. The lipid moiety of brincidofovir is required for in vitro antiviral activity against Ebola virus. *Antivir Res* 2016;125:71–8. <https://doi.org/10.1016/j.antiviral.2015.10.010>.
- [64] Otter JA, Mephams S, Athan B, Mack D, Smith R, Jacobs M, Hopkins S. Terminal decontamination of the Royal Free London's high-level isolation unit after a case of Ebola virus disease using hydrogen peroxide vapor. *Am J Infect Contr* 2016;44(2):233–5. <https://doi.org/10.1016/j.ajic.2015.08.025>. PMID:26521699.
- [65] Dikhit MR, Kumar S, Vijaymahantes Sahoo BR, Mansuri R, Amit A, Yousuf Ansari M, Sahoo GC, Bimal S, Das P. Computational elucidation of potential antigenic CTL epitopes in Ebola virus. *Infection. Genet Evol* 2015;36:369–75. <https://doi.org/10.1016/j.meegid.2015.10.012>. Epub 2015 Oct 14.
- [66] Kiskowski M, Chowell G. Modeling household and community transmission of Ebola virus disease: epidemic growth, spatial dynamics and insights for epidemic control. *Virulence* 2016;7(2):163–73. <https://doi.org/10.1080/21505594.2015.1076613>. 2016.
- [67] Eggers M, Eickmann M, Kowalski K, Zorn J, Reimer K. Povidone-iodine hand wash and hand rub products demonstrated excellent in vitro virucidal efficacy against Ebola virus and modified vaccinia virus Ankara, the new European test virus for enveloped viruses. *BMC Infect Dis* 2015;15:375. <https://doi.org/10.1186/s12879-015-1111-9>. PMID:26381737.
- [68] Steinhubl SR, Marriott MP, Wegerich SW. Remote sensing of vital signs: a wearable, wireless “Band-Aid” sensor with personalized analytics for improved Ebola patient care and worker safety. *Glob Health Sci Pract* 2015;3(3):516–9. <https://doi.org/10.9745/GHSP-D-15-00189>.
- [69] Jacobs M, Aarons E, Bhagani S, Buchanan R, Cropley I, Hopkins S, Lester R, Martin D, Marshall N, Mephams S, Warren S, Rodger A. Post-exposure prophylaxis against Ebola virus disease with experimental antiviral agents: a case-series of health-care workers. *Lancet Infect Dis* 2015;15(11):1300–4. [https://doi.org/10.1016/S1473-3099\(15\)00228-5](https://doi.org/10.1016/S1473-3099(15)00228-5).
- [70] Clay KA, O'Shea MK, Fletcher T, Moore AJ, Burns DS, Craig D, Adam M, Johnston AM, Bailey MS, Gibson CJ. Use of an ultraviolet tracer in simulation training for the clinical management of Ebola virus disease. *Hosp Infect* 2015;91(3):275–7. <https://doi.org/10.1016/j.jhin.2015.07.006>. Epub 2015 Aug 10. PMID:26319591.
- [71] Huttner A, Dayer JA, Yerly S, Combescu C, Auderset F, Desmeules J, Eickmann M, Finckh A, Gonçalves AR, Hooper JW, Kaya G, Krähling V, Kwilas S, Lemaitre B, Matthey A, Silvera P, Becker S, Fast PE, Moorthy V, Kiemy MP, Kaiser L, Siegrist CA. VSV-Ebola Consortium. The effect of dose on the safety and immunogenicity of the VSV-Ebola candidate vaccine: a randomized double-blind, placebo-controlled phase 1/2 trial. *Lancet Infect Dis* 2015;15(10):1156–66. [https://doi.org/10.1016/S1473-3099\(15\)00154-1](https://doi.org/10.1016/S1473-3099(15)00154-1). 2015 Oct.
- [72] Henao-Restrepo AM, Longini IM, Egger M, Dean NE, Edmunds WJ, Camacho A, Carroll MW, Doumbia M, Draguez B, Duraffour S, Enwere G, Grais R, Gunther S, Hossmann S, Kondé MK, Kone S, Kuisma E, Levine MM, Mandal S, Norheim G, Riveros X, Soumah A, Trelle S, Vicari AS, Watson CH, Kéita S, Kiemy MP, Röttingen JA. Efficacy and effectiveness of an rVSV-vectored vaccine expressing Ebola surface glycoprotein: interim results from the Guinea ring vaccination cluster-randomised trial. *Lancet* 2015;386(9996):857–66. [https://doi.org/10.1016/S0140-6736\(15\)61117-5](https://doi.org/10.1016/S0140-6736(15)61117-5).
- [73] Fedson DS, Rordam OM. Treating Ebola patients: a 'bottom up' approach using generic statins and angiotensin receptor blockers. *Int J Infect Dis* 2015;36:80–4. <https://doi.org/10.1016/j.ijid.2015.04.019>. PMID:26143190.
- [74] Broadhurst MJ, Kelly JD, Miller A, Semper A, Bailey D, Groppelli E, Simpson A, Brooks T, Hula S, Nyoni W, Sankoh AB, Kanu S, Jalloh A, Ton Q, Sarchet N, George P, Perkins MD, Wonderly B, Murray M, Pollock NR. ReEBOV Antigen Rapid Test kit for point-of-care and laboratory-based testing for Ebola virus disease: a field validation study. *Lancet* 2015;386(9996):867–74. [https://doi.org/10.1016/S0140-6736\(15\)61042-X](https://doi.org/10.1016/S0140-6736(15)61042-X).
- [75] Wells C, Yamin D, Ndeffo-Mbah ML, Wenzel N, Gaffney SG, Townsend JP, Meyers LA, Fallah M, Nyenswah TG, Altice FL, Atkins KE, Galvani AP. Harnessing case isolation and ring vaccination to control Ebola. *PLoS Neglected Trop Dis* 2015;9(5):e0003794. <https://doi.org/10.1371/journal.pntd.0003794>.
- [76] Althaus CL, Low N, Musa EO, Shuaib F, Gsteiger S. Ebola virus disease outbreak in Nigeria: transmission dynamics and rapid control. *Epidemics* 2015;11:80–4. <https://doi.org/10.1016/j.epidem.2015.03.001>.
- [77] Nyenswah T, Massaquoi M, Gbanya MZ, Fallah M, Amegashie F, Kenta A, Johnson KL, Yahya D, Badini M, Soro L, Pessoa-Silva CL, Roger I, Selvey L, VanderEnde K, Murphy M, Cooley LA, Olsen SJ, Christie A, Vertefeuille J, Navin T, McElroy P, Park BJ, Esswein E, Fagan R, Mahoney F. Centers for Disease Control and Prevention (CDC). Initiation of a ring approach to infection prevention and control at non-ebola health care facilities — Liberia, January–February 2015. *Morb Mortal Wkly Rep* 2015;64(18):505–8.
- [78] Khan MA, Hossain MU, Rakib-Uz-Zaman SM, Morshed MN. Epitope-based peptide vaccine design and target site depiction against Ebola viruses: an immunoinformatics study. *Scand J Immunol* 2015;82(1):25–34.
- [79] Fähnrich C, Denecke K, Adeyoye OO, Benzler J, Claus H, Kirchner G, Mall S, Richter

- R, Schapranow MP, Schwarz N, Tom-Abu D, Uflacker M, Poggensee G, Krause G. Surveillance and outbreak response management system (SORMAS) to support the control of the Ebola virus disease outbreak in West Africa. *Euro Surveill* 2015;20(12). 2015 Mar 26;pii: 21071.
- [80] Chin AW, Perera RA, Guan Y, Halfmann P, Kawaoka Y, Peiris M, Poon LL. Pseudoparticle neutralization assay for detecting Ebola- neutralizing antibodies in biosafety level 2 settings. *Clin Chem* 2015;61(6):885–6.
- [81] Regules JA, Beigel JH, Paolino KM, Voell J, Castellano AR, Hu Z, Muñoz P, Moon JE, Ruck RC, Bennett JW, Twomey PS, Gutiérrez RL, Remick SA, Hack HR, Wisniewski ML, Josley MD, Kwilas SA, Van Deusen N, Mbaya OT, Zhou Y, Stanley DA, Jing W, Smith KS, Shi M, Ledgerwood JE, Graham BS, Sullivan NJ, Jagodzinski LL, Peel SA, Alimonti JB, Hooper JW, Silvera PM, Martin BK, Monath TP, Ramsey WJ, Link CJ, Lane HC, Michael NL, Davey Jr. RT, Thomas SJ. rVSVΔG-ZEBOV-GP Study Group. A recombinant vesicular stomatitis virus Ebola vaccine. *N Engl J Med* 2017;376(4):330–41.
- [82] Agnandji ST, Huttner A, Zinser ME, Njuguna P, Dahlke C, Fernandes JF, Yerly S, Dayer JA, Kraehling V, Kasonta R, Adegnik J, Altfeld M, Auderset F, Bache EB, Biedenkopf N, Borregaard S, Brosnahan JS, Burrow R, Combesure C, Desmeules J, Eickmann M, Fehling SK, Finckh A, Goncalves AR, Grobusch MP, Hooper J, Jambrecina A, Kabwende AL, Kaya G, Kimani D, Lell B, Lemaître B, Lohse AW, Massinga-Loembe M, Matthey A, Mordmüller B, Nolting A, Ogwang C, Ramharter M, Schmidt-Chanasis J, Schmiedel S, Silvera P, Stahl FR, Staines HM, Strecker T, Stubbe HC, Tsfoa B, Zaki S, Fast P, Moorthy V, Kaiser L, Krishna S, Becker S, Kieny MP, Bejon P, Kremsner PG, Addo MM, Siegrist CA. Phase 1 trials of rVSV Ebola vaccine in Africa and Europe. *N Engl J Med* 2016;374(17):1647–60. <https://doi.org/10.1056/NEJMoa1502924>.
- [83] Zhu FC, Hou LH, Li JX, Wu SP, Liu P, Zhang GR, Hu YM, Meng FY, Xu JJ, Tang R, Zhang JL, Wang WJ, Duan L, Chu K, Liang Q, Hu JL, Luo L, Zhu T, Wang JZ, Chen W. Safety and immunogenicity of a novel recombinant adenovirus type-5 vector-based Ebola vaccine in healthy adults in China: preliminary report of a randomised, double-blind, placebo-controlled, phase 1 trial. *Lancet* 2015;385(9984):2272–9. [https://doi.org/10.1016/S0140-6736\(15\)60553-0](https://doi.org/10.1016/S0140-6736(15)60553-0).
- [84] Weitz JS, Dushoff J. Modeling post-death transmission of Ebola: challenges for inference and opportunities for control. *Sci Rep* 2015;5:8751. <https://doi.org/10.1038/srep08751>.
- [85] Sakurai Y, Kolokoltsov AA, Chen CC, Tidwell MW, Bauta WE, Klugbauer N, Grimm C, Wahl-Schott C, Biel M, Davey RA. Ebola virus. Two-pore channels control Ebola virus host cell entry and are drug targets for disease treatment. *Science* 2015;347(6225):995–8. <https://doi.org/10.1126/science.1258758>.
- [86] Ewer K, Rampling T, Venkatraman N, Bowyer G, Wright D, Lambie T, Imoukhuede EB, Payne R, Fehling SK, Strecker T, Biedenkopf N, Kröhling V, Tully CM, Edwards NJ, Bentley EM, Samuel D, Labbi G, Jin J, Gibani M, Minhinnick A, Wilkie M, Poulton I, Lella N, Roberts R, Hartnell F, Bliss C, Sierra-Davidson K, Powlson J, Berrie E, Tedder R, Roman F, De Ryck I, Nicosia A, Sullivan NJ, Stanley DA, Mbaya OT, Ledgerwood JE, Schwartz RM, Siani L, Colloca S, Folgori A, Di Marco S, Cortese R, Wright E, Becker S, Graham BS, Koup RA, Levine MM, Volkman A, Chaplin P, Pollard AJ, Draper SJ, Ballou WR, Lawrie A, Gilbert SC, Hill AV. A monoavalent chimpanzee adenovirus Ebola vaccine boosted with MVA. *N Engl J Med* 2016;374(17):1635–46. <https://doi.org/10.1056/NEJMoa1411627>.
- [87] Johnson JC, Martinez O, Honko AN, Hensley LE, Olinger GG, Basler CF. Pyridinyl imidazole inhibitors of p38 MAP kinase impair viral entry and reduce cytokine induction by Zaire Ebola virus in human dendritic cells. *Antivir Res* 2014;107:102–9. <https://doi.org/10.1016/j.antiviral.2014.04.014>. 2014 Jul.
- [88] Setlur AS, Naik SY, Skariyachan S. Herbal lead as ideal bioactive compounds against probable drug targets of Ebola virus in comparison with known chemical analogue: a computational drug discovery perspective, interdisciplinary sciences. *Computational Life Sci* 2017;9(2):254–77.
- [89] Srivastava PN, Jain R, Dubey SD, Bhatnagar S, Ahmad N. International journal of peptide research and therapeutics vol. 22. 2016. p. 119–33. (1).
- [90] Raj U, Varadwaj PK. Flavonoids as multi-target inhibitors for proteins associated with Ebola virus: in silico discovery using virtual screening and molecular docking studies. *Interdiscipl Sci Comput Life Sci* 2016;8(2):132–41.
- [91] Krähling V, Becker D, Rohde C, Eickmann M, Eroğlu Y, Herwig A, Kerber R, Kowalski K, Vergara-Alert J, Becker S. European Mobile Laboratory consortium. Development of an antibody capture ELISA using inactivated Ebola Zaire Makona virus. *Med Microbiol Immunol* 2016;205(2):173–83.
- [92] Balmith M, Soliman MES. VP40 of the Ebola virus as a target for EboV therapy: comprehensive conformational and inhibitor binding landscape from accelerated molecular dynamics. *Cell Biochem Biophys* 2017;75(1):65–78.
- [93] Oany AR, Sharmin T, Chowdhury AS, Jyoti TP, Hasan MA. Highly conserved regions in Ebola virus RNA dependent RNA polymerase may be act as a universal novel peptide vaccine target: a computational approach. *In Silico Pharmacol* 2015;3(1):7. <https://doi.org/10.1186/s40203-015-0011-4>.
- [94] Qiu S, Leung A2, Bo Y1, Kozak RA2, Anand SP1, Warkentin C1, Salambanga FDR1, Cui J1, Kobinger G2, Kobasa D3, Côté M4. Ebola virus requires phosphatidylinositol (3,5) biphosphate production for efficient viral entry. *Virology* 2017;513:17–28. <https://doi.org/10.1016/j.virol.2017.09.028>.
- [95] Bhattacharyya S, Warfield KL, Ruthel G, Bavari S, Aman MJ, Hope TJ. Ebola virus uses clathrin-mediated endocytosis as an entry pathway. *Virology* 2010;401(1):18–28. <https://doi.org/10.1016/j.virol.2010.02.015>.
- [96] Wang Y, Liu Z, Dai Q. A highly immunogenic fragment derived from Zaire Ebola virus glycoprotein elicits effective neutralizing antibody. *Virus Res* 2014;30(189):254–61. <https://doi.org/10.1016/j.virusres.2014.06.001>. 2014 Aug.
- [97] Trunschke M, Conrad D, Enterlein S, Olejnik J, Brauburger K, Mühlberger E. The L-VP35 and L-L interaction domains reside in the amino terminus of the Ebola virus L protein and are potential targets for antivirals. *Virology* 2013;441(2):135–45. <https://doi.org/10.1016/j.virol.2013.03.013>.
- [98] Lee JE, Kuehne A, Abelson DM, Fusco ML, Hart MK, Saphire Erica Ollmann. Complex of a protective antibody with its Ebola virus GP peptide epitope: unusual features of a V<sub>L</sub>x-light chain. *J Mol Biol* 2008;375(1):202–16.
- [99] He F, Melén K, Maljanen S, Lundberg R, Jiang M, Österlund P, Kakkola L, Julkunen I. Ebolavirus protein VP24 interferes with innate immune responses by inhibiting interferon-λ1 gene expression. *Virology* 2017;509:23–34. <https://doi.org/10.1016/j.virol.2017.06.002>.
- [100] Oluwagbemi O, Oluwagbemi F, Abimbola O. Ebinformatics: Ebola fuzzy informatics systems on the diagnosis, prediction and recommendation of appropriate treatments for Ebola virus disease (EVD). *Informatics in Medicine Unlocked* 2016;2(2016):12–37.
- [101] Su Z, Wu C, Shi L, Leung DW, Chiu W, Amarasinghe GK. Electron cryo-microscopy structure of Ebola virus nucleoprotein reveals a mechanism for nucleocapsid-like assembly. *Cell* 2018;172(5):966–78.
- [102] Kaletsky RL. Host cell factors involved in Ebola virus entry and spread, a PhD dissertation in cell and molecular biology, submitted to the university of Pennsylvania. 2009.
- [103] Brangel P, Sobarzo A, Parolo C, Miller BS, Howes PD, Gelkop S, Lutwama JJ, Dye JM, McKendry RA, Lobel L, Stevens MM. A serological point-of-care test for the detection of IgG antibodies against Ebola virus in human survivors. *ACS Nano* 2018;12(1):63–73. <https://doi.org/10.1021/acsnano.7b07021>.
- [104] Ponomarenko J, Kerrie Vaughan K, Paul S. Ebola: an analysis of immunity at the molecular level. 2015 international workshop on artificial immune systems (AIS), date of conference: 17–18 July 2015 held at Taormina, Italy. 2015.
- [105] Abu-haraz AH, Abd-elrahman2 Khoubieb Ali, Ibrahim2 Mojahid Salah, Hussien2 Waleed Hassan, Mohammed1 Mohammed Siddiq, Badawi1 Marwan Mustafa, Saleh1 Mohamed Ahmed. Multi epitope peptide vaccine prediction against Sudan Ebola virus using immuno-informatics approaches. *Adv Tech Biol Med* 2017;5:203.
- [106] Ayuba G, Waheed Y, Najmi MH. Prediction and conservancy analysis of promiscuous T-cell binding epitopes of Ebola virus L protein: an in silico approach. *Asian Pac J Trop Dis* 2016;6(3):169–73.
- [107] Grifoni A, LoPresti Alessandra, Giovanetti Marta, Montesano Carla, Amicosante Massimo, Colizzi Vittorio, Lai Alessia, Zehender Gianguglielmo, Cella Eleonora, Angeletti Silvia, Cicozzi Massimo. Genetic diversity in Ebola virus: phylogenetic and in silico structural studies of Ebola viral proteins. *Asian Pac J Trop Med* 2016;9(4):337–43.
- [108] Khurana S, Fuentes S, Coyle EM, Ravichandran S, Davey Jr. RT, Beigel JH. Human antibody repertoire after VSV-Ebola vaccination identifies novel targets and virus-neutralizing IgM antibodies. *Nat Med* 2016;22(12):1439–47. <https://doi.org/10.1038/nm.4201>. 2016 Dec.
- [109] Yasmin T, Nabi AH. B and T Cell epitope-based peptides predicted from evolutionarily conserved and whole protein sequences of Ebola virus as vaccine targets. *Scand J Immunol* 2016;83(5):321–37. <https://doi.org/10.1111/sji.12425>.
- [110] Srivastava PN, Jain R, Dubey SD, Bhatnagar S, Ahmad N. Prediction of epitope-based peptides for vaccine development from coat proteins GP2 and VP24 of Ebola virus using immunoinformatics. *Int J Pept Res Therapeut* 2016;22(1):119–33.
- [111] Radostitzky SR, Warfield Kelly L, Chi Xiaoli, Dong Lian, Kota Krishna, Bradfute Steven B, Jacqueline D Gearhart, Retterer Cary, Kranzusch Philip J, Misasi John N, Hogenbirk Marc A, Wahl-Jensen Victoria, Volchkov Viktor E, Cunningham James M, Jahrling Peter B, Aman M Javad, Bavari1 Sina, Farzan Michael, Kuhn Jens H. Ebolavirus Δ-peptide immunoadhesins inhibit marburgvirus and ebolavirus cell entry. *J Virol* 2011;85(17):8502–13.
- [112] Mirza UM, Ikram Nazia. Integrated computational approach for virtual hit identification against Ebola viral proteins VP35 and VP40. *Int. J. Mol. Sci.* 2016;17(11):1748. <https://doi.org/10.3390/ijms17111748>. 2016 Nov.
- [113] Edgar Robert C. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res* 2004;32(5):1792–7.
- [114] Zhang Zheng, Schwartz Scott, Wagner Lukas, Miller Webb. A greedy algorithm for aligning DNA sequences. *J Comput Biol* 2000;7(1–2):203–14. 2000.
- [115] Gouy M, Guindon S, Gascuel O. SeaView version 4: a multiplatform graphical user interface for sequence alignment and phylogenetic tree building. 2010.
- [116] Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, Valentin F, Wallace IM, Wilm A, Lopez R, Thompson JD, Gibson TJ, Higgins DG. Clustal W and clustal X version 2.0. *Bioinformatics* 2007;23:2947–8.
- [117] Stamatakis Alexandros. Raxml-vi-hpc: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 2006;22(21):2688–90.
- [118] Page RDM. TREEVIEW: an application to display phylogenetic trees on personal computers. *Comput Appl Biosci* 1996;12:357–8.
- [119] Letunic and Bork. Nucleic Acids Res 2016. <https://doi.org/10.1093/nar/gkw290>.
- [120] Page Roderic DM. Visualizing phylogenetic trees using TreeView. 2002. Volume00, Issue1, January 2003 Pages 6.2.1–6.2.15.
- [121] Baize S, Pannetier Delphine, Oestereich Lisa, Rieger Toni, Koivogui Lamine, Magassouba N’Faly, Soropogui Barré, Saliou Sow Mamadou, Keita akoba, De Clerck Hilde, Tiffany Amanda, Dominguez Gemma. Emergence of Zaire Ebola virus disease in Guinea. *N Engl J Med* 2014;371:1418–25. <https://doi.org/10.1056/NEJMoa1404505>.
- [122] Wauquier N, Becquart P, Padilla C, Baize S, Leroy EM. Human fatal Zaire Ebola virus infection is associated with an aberrant innate immunity and with massive lymphocyte apoptosis. *PLoS Neglected Trop Dis* 2010;4(10):e8377. <https://doi.org/10.1371/journal.pntd.0000837>.
- [123] Harris JW, Stocker H. Maximum likelihood method. §21.10.4 in handbook of mathematics and computational science. New York: Springer-Verlag; 1998. 824,

- 1998.
- [124] Efron B, Tibshirani R. An introduction to the bootstrap. Boca Raton, FL: Chapman & Hall/CRC; 1993. ISBN 0-412-04231-2[Software].
- [125] Weisstein Eric W. "Bootstrap methods." from MathWorld—a wolfram web resource. <http://mathworld.wolfram.com/BootstrapMethods.html>.
- [126] Varian H. Bootstrap tutorial. *Math J* 2005;9:768–75. 2005. Extra [127].
- [127] Purvis A, Katzourakis A, Agapow PM. Evaluating phylogenetic tree shape: two modifications to Fusco & Cronk's method. *J Theor Biol* 2002;214(1):99–103.
- [128] Chakerian John, Holmes Susan. Computational tools for evaluating phylogenetic and hierarchical clustering trees, by Chakerian and holmes, 2010. 2010. arXiv:1006.1015 [stat.AP].
- [129] Xia XI, Xie Z. DAMBE: software package for data analysis in molecular biology and evolution. *J Hered* 2001;92(4):371–3. 2001 Jul-Aug.
- [130] DAMBE 5. A comprehensive software package for data analysis in molecular biology and evolution. *Mol Biol Evol* 2013 Jul;30(7):1720–8. <https://doi.org/10.1093/molbev/mst064>. Published online 2013 Apr 5.
- [131] Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O. New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. *Syst Biol* 2010;59(3):307–21. <https://doi.org/10.1093/sysbio/syq010>. Epub 2010 Mar 29.
- [132] Kumar S, Tamura K, Nei M. MEGA3: integrated software for molecular evolutionary genetics analysis and sequence alignment. *Briefings Bioinf* 2004;5(2):150–63.
- [133] Schmidt HA, Strimmer K, Vingron M, von Haeseler A. TREE-PUZZLE: maximum likelihood phylogenetic analysis using quartets and parallel computing. *Bioinformatics* 2002;18(3):502–4.
- [134] Felsenstein J. Confidence limits on phylogenies: An approach using the bootstrap. *Evolution* 1985;vol. 39(4):783–91. <https://doi.org/10.1111/j.1558-5646.1985.tb00420.x>. 1985 Jul.
- [135] Olsen GJ, Matsuda H, Hagstrom R, Overbeek R. fastDNAm1: a tool for construction of phylogenetic trees of DNA sequences using maximum likelihood. *Comput Appl Biosci* 1994;10(1):41–8.
- [136] Kumar S, Tamura K, Nei M. MEGA: molecular evolutionary genetics analysis software for microcomputers. *Bioinformatics* 1994;189–91.
- [137] Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony method. *Mol Biol Evol* 2011;28(10):2731–9 <https://doi.org/10.1093/molbev/msr121>.
- [138] Yang Z. PAML 4: phylogenetic analysis by maximum likelihood. *Mol Biol Evol* 2007;24(8):1586–91.
- [139] Guindon S, Delsuc F, Dufayard JF, Gascuel O. Estimating maximum likelihood phylogenies with PhyML. *Meth Mol Biol* 2009;537:113–37. [https://doi.org/10.1007/978-1-59745-251-9\\_6](https://doi.org/10.1007/978-1-59745-251-9_6).
- [140] Strimmer K, von Haeseler A. Quartet puzzling: a Quartet-maximum likelihood method for reconstructing tree topologies. *Mol Biol Evol* 1996;13(7). 1 September 1996, Pages 964 <https://doi.org/10.1093/oxfordjournals.molbev.a025664>.
- [141] Larget B, Simon DL. Markov chain Monte Carlo algorithms for the bayesian analysis of phylogenetic trees. *Mol Biol Evol* 1999;16(6):750–9.
- [142] Drummond AJ, Rambaut A. BEAST: bayesian evolutionary analysis by sampling trees. *BMC Evol Biol* 2007;8(7):214. 2007 Nov.
- [143] Xia X. DAMBE5: a comprehensive software package for data analysis in molecular biology and evolution. *Mol Biol Evol* 2013;30(7):1720–8. <https://doi.org/10.1093/molbev/mst064>.
- [144] Matsen FA, Kodner RB, Armbrust EV. pplacer: linear time maximum-likelihood and Bayesian phylogenetic placement of sequences onto a fixed reference tree. *BMC Bioinf* 2010;11:538. <https://doi.org/10.1186/1471-2105-11-538>.
- [145] Than C, Ruths D, Nakhleh L. PhyloNet: a software package for analyzing and reconstructing reticulate evolutionary relationships. *BMC Bioinf* 2008;9:322. <https://doi.org/10.1186/1471-2105-9-322>.
- [146] Trifinopoulos J, Nguyen LT, von Haeseler A, Minh BQ. W-IQ-TREE: a fast online phylogenetic tool for maximum likelihood analysis. *Nucleic Acids Res* 2016;44(W1):W232–5. <https://doi.org/10.1093/nar/gkw256>.
- [147] Felsenstein J. Inferring phylogenies from protein sequences by parsimony, distance, and likelihood methods. *Meth Enzymol* 1996;1996(266):418–27.
- [148] Rohlf FJ, Wooten MC. Evaluation of the restricted maximum-likelihood method for estimating phylogenetic trees using simulated allele-frequency data. *Evolution* 1988;42(3):581–95. <https://doi.org/10.1111/j.1558-5646.1988.tb04162.x>. 1988 May.
- [149] Felsenstein J. Numerical methods for inferring evolutionary trees. *Q Rev Biol* 1982;57(4):379–404.
- [150] Chang JM, Di Tommaso P, Notredame C. TCS: a new multiple sequence alignment reliability measure to estimate alignment accuracy and improve phylogenetic tree reconstruction. *Mol Biol Evol* 2014;31(6):1625–37. <https://doi.org/10.1093/molbev/msu117>.
- [151] Yang Ziheng. Evaluation of several methods for estimating phylogenetic trees when substitution rates differ over nucleotide sites. *J Mol Evol* June 1995;40(6):689–97 <https://link.springer.com/article/10.1007/BF00160518>.
- [152] Ignacio Requeno José, Colom José Manuel. Evaluation of properties over phylogenetic trees using stochastic logics. *BMC Bioinf* 2016;2016(17):235. <https://doi.org/10.1186/s12859-016-1077-7>. Published online 2016 Jun 14.
- [153] Rosemary M, McCloskey, Richard H, Liang P, Richard Harrigan, Brumme Zabrina L, Poo Art FY. An evaluation of phylogenetic methods for reconstructing transmitted HIV variants using longitudinal clonal HIV sequence data. *J Virol* 2014;88(11):6181–94. <https://doi.org/10.1128/JVI.00483-14>. 2014 Jun.
- [154] Andypurvis1, Ariskatzourakis, Paul-Michaelagapow. Evaluating phylogenetic tree shape: two modifications to Fusco & Cronk's method. *J Theor Biol* 2002;214(1):99–103. 7 January 2002, Pages.
- [155] Chakerian John, Holmes Susan. Computational tools for evaluating phylogenetic and hierarchical clustering trees. 2010. arXiv:1006.1015 [stat.AP].
- [156] Enosawa Ryosuke, Mutoh Atsuko, Inuzuka Nobuhiro. A phyletic model for evaluating phylogenetic tree estimation, Consumer Electronics (GCCE). 2015 IEEE 4th Global conference on, 27–30 Oct. 2015. Osaka, Japan: IEEE; 2015 <http://ieeexplore.ieee.org/document/7398622/>.
- [157] Horiike Tokumasa. An introduction to molecular phylogenetic analysis. *Rev Agric Sci* 2016;4:36–45. <https://doi.org/10.7831/ras.4.36>. 2016 <http://www.agrsci.jp/ras/article/view/21/43>.
- [158] Felsenstein J. PHYLIP - phylogeny inference package (version 3.2). *Cladistics* 1989;5:164–6 <http://evolution.genetics.washington.edu/phylip.html>.
- [159] Kumar S, Stecher G, Li M, Knyaz C, Tamura K. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Mol Biol Evol* 2018;35:1547–9.
- [160] ETE 3: Reconstruction, analysis and visualization of phylogenomic data. Jaime Huerta-Cepas, Francois Serra and Peer Bork *Mol Biol Evol* 2016. <https://doi.org/10.1093/molbev/msw046> <http://etetoolkit.org/treeview/>.
- [161] Webb CO, Ackerly DD, Kembel SW. Phylocom: software for the analysis of phylogenetic community structure and trait evolution. *Bioinformatics* 2008;24(18):2098–100. <https://doi.org/10.1093/bioinformatics/btn358>. Epub 2008 Aug 4.